





BABEŞ-BOLYAIUNIVERSITY FROM CLUJ-NAPOCA FACULTY OF CHEMISTRY AND CHEMICAL ENGINEERING DEPARTMENT OF CHEMICAL ENGINEERING SCIENTIFIC RESEARCH CENTER IN PHYSICAL CHEMISTRY

ABSTRACT OF DOCTORAL THESIS

PREPARATION AND CHARACTERIZATION OF NANOHYDROXYAPATITES DOPED WITH VARIOUS BIOLOGICAL ACTIVE COMPOUNDS WITH BIOMEDICAL APPLICATIONS

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| INTRODUCTION |
|--|
| 1. Enhancement of bone consolidation using highfrequency pulsed electromagnetic short-waves and titanium implants coated with biomimetic compositeembedded into |
| PLA matrix: in vivo evaluation. |
| Abstract |
| 1.1. Introduction 2 |
| 1.2. Experimental section |
| 1.2.1 Materials and sample preparation |
| 1.2.2. Titanium (Ti) rods |
| 1.2.3. Preparation of ms-HAP/COL nanomaterial |
| 1.2.4. HAPc coating of Ti implants 7 |
| 1.3. Characterization methods of composites |
| 1.3.1. X-ray diffraction (XRD) |
| 1.3.2. High-resolution-transmission electron microscope (HR_TEM) |
| 1.3.3. Scanning electron microscope (SEM) |
| 1.3.4. Atomic force microscope (AFM) 10 |
| 1.3.5. Study protocol |
| 1.3.6. Surgical procedures |
| 1.3.7. Bone markers determination |
| 1.3.8. Histological assessment. Hematoxylin andeosin (H&E) staining |
| 1.3.9. Micro-computed tomography |
| 1.3.10, Three points bending test. |
| 1.3.11. Statistical analysis |
| 1.4. Results |
| 1.4.1.XRD 13 |
| 142 Morphology and surface structure |
| 143 Animal studies |
| 144 Rome markers concentration |
| 1.4.5 Direc-commuted tomography 15 |
| 1.4.6 Three points barding tests |
| 1.4.0. Three points behaviors in the second se |
| 15 Disussions 21 |
| 1.5. Discussions |
| 1.7. Defermance 20 |
| 1.7. References |
| 2. Biocompatibility of titanium implants coated with biocomposite in a rat model of |
| femoral fracture |
| Abstract |
| 2.1.Introduction |
| 2.2. Results and discussion |
| 2.2.1. Surface morphology of HAPc biocomposite coated Ti implants |
| 2.2.2. Bone markers: alkaline phosphatase and osteocalcin results |
| 2.2.3. Micro-CT results |
| 2.2.4. Histological results |
| 2.3. Conclusions |
| 2.4. Experimental section 48 |
| 2.4.1. Materials, sample preparation and characterization |
| 2.4.2. Study protocol |
| 2.4.3. Surgical procedures 49 |
| 2.4.4. Alkaline phosphatase and osteocalcin. |
| 2.4.5 Micro-CT 50 |
| 2.4.6 Histological assessment 50 |
| 2.4.7 Statistical analysis 50 |
| References 51 |
| J1 |
| 3. Behavior of multisubstituted hydroxyapatites in water and simulated body fluid |
| |

CONTENTS

| 3.1. Introduction | 7 |
|---|----------------|
| 3.2. Results and discussion | 3 |
| 3.2.1. Structural and morphological characterization |) |
| 3.2.2. Behavior in water | 2 |
| <i>3.2.3. Behavior in SBF</i> 63 | 3 |
| 3.3. Conclusions | 5 |
| 3.4. Experimental section | 5 |
| 3.4.1. Samples preparation |) |
| 3.4.2. Samples characterization |) 7 |
| 3.5 Pafarances | 7 |
| 5.5. Keleenees | , |
| 4. Higuchi model applied to ions release rate from hydroxyapatites | 3 |
| Abstract | 3 |
| 4.1. Introduction | 1 |
| 4.2. Results and discussion | 5 |
| 4.3. Conclusions | 3 |
| 4.4. Experimental section | 3 |
| 4.5. References | ŧ |
| | |
| 5. Antibacterial activity of silver nanoparticles obtained by co-reduction with sodium |) |
| citrate and fannic acid. |) |
| ADSITACT | ź |
| 5.2 Popults and discussion 91 | l |
| 5.2. Results and discussion. | 2 |
| 5.2.1. 0 V-V15 spectra | 1 |
| 5.2.2. Antimicrohial activity | 7 |
| 53 Conclusions |) |
| 5.4. Experimental section. |) |
| 5.4.1. Synthesis of silver nanoparticles (AeNP |) |
| 5.4.2. Measurements and instrumentation. |)() |
| 5.5. References |)1 |
| | 17 |
| 6. Interaction of silver nanoparticles with vancomycin: an UV-VIS study |)7 |
| Abstract |)7 |
| 6.1. Introduction |)8 |
| 6.2. Conclusions | 12 |
| 6.4 Experimental section 11 | 13 |
| 6.4.1 Synthesis of silver nanoparticles | 13 |
| 6.4.2 Interactionea AoNP cu vancomicina | 14 |
| 6.5. References. | 14 |
| | |
| 7. Biomimetic nanocomposite structures designed for the coating of orthopedic implants: | 10 |
| AFM investigation. | 19 |
| Abstract | 20 |
| 7.1. Introduction | 20 |
| 7.2. Results and discussion. | 33 |
| 7.3. Conclusions | 34 |
| 7.4.1. Titanian control of the management of the section 13 | 34 |
| 7.4.2 Propagation of the coating dispersions | 34 |
| 7.4.2. Freparation of the coating aispersions | 35 |
| 7.4.4 Y ray diffraction (YRD) 13 | 35 |
| 7.4.5 AFM microscony 13 | 35 |
| 7.5. References | 36 |
| 8. General conclusions | 42 |
| 9. Selective references | 1 6 |

| 10. Original research activity results | 165 |
|---|-----|
| 10.1. List of published papers | 165 |
| 10.2. List of communications at conferences and symposia | 167 |
| 10.3. List of scientific grants | 169 |
| 10.4. Member in the Center of Scientific Research in Physical Chemistry | 170 |

Keywords

stoichiometric hydroxyapatite; multisubstituted hydroxyapatite, magnesium, zinc, silicon, strontium; silver nanoparticles; vancomycin; collagen; PLA; XRD; SEM-EDX; AFM; Ti implants; Ti implants coated with biomimetic composites; biocompatibility; antimicrobial activity; *in vivo* rat model; femoral fracture; ALP; OCN; histological analysis; micro-CT; mechanical properties; new bone formation; bone fracture healing.

Introduction

Bone replacement and bone repair are the most controversial methods of bone regeneration in orthopedic surgery. For medical applications, it is necessary to develop materials that mimic chemical composition and structure of natural bone and favour faster healing with minimal side effects. The bone of the human body is one of the largest organs that perform the functions of movement, support, mineral intake and protection. It also has the ability to reshape and self- healing that means absorption of old or of damaged bone tissue occurring at the same interface where osteoblasts produce new healthy bone to replace resorbed bone. However, in the critical cases when the bone is severely damaged, self-healing is not enough. Therefore, biomaterials capable of substituting or regenerating the affected bone tissue need to be developed.

Basically, the bone tissue engineering seems to solve these problems utilizing a combination of multidisciplinary approaches to improve or replace damaged bone tissue. In recent years, due to the development of tissue engineering technology, bone tissue engineering has become an optimistic approach for repairing bone fractures and bone defects. Biomaterials for bone substitutes and for coating of metallic implants or as scaffolds in cell culture were developed in this thesis [1-14] and play a crucial role in bone tissue engineering. Their purpose is to mimic the composition, structure and function of the natural bone and provide a three-dimensional (3D) environment to have adequate physical properties for bone repair promoting the adhesion, proliferation, and differentiation of osteoblast cells *in vivo*[5,6].

An ideal nanostructured biomaterial, also named biomedical material should be biodegradable, bioactive, biocompatible and osteoconductive or especially osteoinductive *in vivo*, proving the new bone formation and growth. The goal of this work was to design, prepare and characterize synthetic nanohydroxiapatites, nano HAPs, which are important components of natural bone [1-14], doped with various biological active compounds, such as collagen (COL, another essential component in bone), and drugs (e.g. antibiotics, like vancomycin and silver nanoparticles:

AgNPs to protect against infections [6, 8] jointly able to develop innovative nanomaterials with enhanced biological functions for bone substitutes and for coating of metallic implants with potential biomedical applications, for biological bone repair, enhanced bone fracture healing and bone regeneration [7, 9, 12, 14].

Due to the various needs of synthetic nanostructured biomaterials, like nanohydroxiapatites multi-doped with essential physiological elements, such as Mg, Zn and Si, resulting multi-substituted hydroxyapatites, noted ms-HAP or HAPc, were also developed in this work to enhance the bone regeneration [1-14].

The multidisciplinary approaches developed in this work can be applied for bone regeneration and will make important steps in the near future concerning the exploitation of novel biomaterials and new strategies regarding the integration of nanotechnology, in stem cell science for bone tissue regeneration helping the population at risk, such as patients with osteoporosis.

The design, synthesis, and physico-chemical characterization, as well as their biological behavior were carefully investigated in this doctoral thesis under the leadership of Univ. Prof. Dr. Maria Tomoaia-Cotisel, Director of the Research Center in Physical Chemistry, CECHIF, at Babeş-Bolyai University of Cluj-Napoca. The results obtained in this doctoral thesis are part of significant achievements obtained in the last decade in the CECHIF center, including *in vivo*studies using *femoral fracture rat model*.

Chapter 1 [7] aims to investigate *nanostructured biomaterials* made within hedoctoral research, *nanostructured biomimetic composite* materials, containing three components: HAPc nanoparticles functionalized with COL crown (core / shell nanoparticles), incorporated in polylactic acid, PLA, resulting in porous coatings HAPc-COL @ PLA on Ti implants. Finally, these composites were coated with self-assembled COL fibers, resulting in HAPc-COL@PLA/COL biomimetic materials [7]. The addition of collagen to the porous HAPc-COL@PLA coating material increased the mechanical strength of the composite, causing a reduction in its porosity. The materials were obtained by the self-assembling dip coating method on Ti implants. The materials were characterized by top methods and techniques, namely XRD, TEM, SEM, EDX, AFM (Figure 1), and methods for determining the mechanical properties (Figure 5, Table 1) of the implants used. The materials were studied *in vivo*, on a *model of femoral fractures in rats* which represents a *premiere in the international scientific community*.



Figure 2. XRD patterns for the lyophilized ms-HAP/COL (A) powder and for the HAPc composite (i.e., the ms-HAP/COL@PLA/COL coating) on the Ti surface(B); HR-TEM images (C, D) : shapes of HAP/COL nanoparticle; SEM image (E) and EDX spectrum for the same area (F) for HAP/COL; SEM images (G, H) of the HAPc coating on Ti surface.

Figure 5 Force vs displacement curves for the three points bending tests performed on the explanted femur of rats: (A) at 2 weeks post-implantation and (B) at 8 weeks post-implantation

Table 1 Measured values of the breaking force and the corresponding displacement during the three points bending tests.

| Grou | р | CG | PESW | HAPc | HAPc+PE SW |
|----------------------------------|---------|---------------|-------------|---------------|---------------|
| Ultimate force[N] | 2 weeks | 20 ± 5 | 34 ± 4 | 30 ± 4 | 40 ± 5 |
| | 8 weeks | 95 ± 9 | 124 ± 6 | 121 ± 8 | 139 ± 8 |
| Ultimate displacement [mm] | 2 weeks | 1.5 ± 0.3 | 0.6± 0.2 | 1.0 ± 0.3 | 0.7 ± 0.1 |
| | 8 weeks | 0.6 ± 0.1 | 0.5 ± 0.1 | 0.5 ± 0.2 | 0.5 ± 0.1 |

Abbreviations: CG = control group; PESW = pulsed electromagnetic short-waves; HAPc = titanium implants coated with multisubstituted hydroxyapatite and collagen; HAPc+PESW = titanium implants coated with multisubstituted hydroxyapatite and collagen and pulsed electromagnetic short-waves; N = newton; mm = milimeter.

The results regarding an *in vivo* evaluation of the enhancement of bone consolidation using highfrequencypulsed electromagnetic short-waves and titanium implants coated with biomimetic composite embedded into PLA matrix are presented. This research demonstrates the importance of biomimetic coatings deposited on titanium implants in the rapid healing of femoral fractures in the rat model. The multidisciplinary research carried out includes the determination of bone markers: alkaline phosphatase and osteocalcin, at different implantation times in animals (at 2, 4 and 8 weeks, respectively); the technique of micro computertomography (micro-CT and the histological analysis) are used, and the fast healing of the femoral fractures in the rat is highlighted.

Chapter 2 [9] reports on *Biocompatibility of titanium implants coated with biocomposite in a rat model of femoral fracture* and demonstrates *in vivo*the osseointegration of the biomimetic coating on Ti implants with the host natural bone. Measurements of bone marker biophysics: alkaline phosphatase and osteocalcin (Figure 2, Table 1), as well as micro-CT (Table 2) and histological analysis (Figure 3). They demonstrateosseointegration (Table 2), the formation of trabecular bone and compact bone - facilitated by the biominetic structures designed and developed in this doctoral thesis.

Figure 2. Bone markers, alkaline phosphatase (A) and osteocalcin (B), serum concentration at zero time (initially), two- and eight-weeks post-surgery; *statistically significant p<0.05; **statistically significant p<0.01; ***statistically significant p<0.001; ***statistically significant p<0.001

Table 1. Bone markers, alkaline phosphatase (ALP) and osteocalcin (OCN), serum concentration at initial (0 weeks), two- and eight-weeks post-operatively; *statistically significant p<0.05; **statistically significant p<0.01; ***statistically significant p<0.001; ***statistically significant p<0.001

| Rat group |) | CG | HAPc |
|-----------|---------|---------------------|---------------------|
| ALP | 0 weeks | 100 ± 13 | 102 ±6 |
| (%) | 2 weeks | 152 ±14**** | $173 \pm 10^{****}$ |
| | 8 weeks | 100 ± 7 | 89 ± 8 |
| OCN | 0 weeks | 100 ± 15 | 104 ± 13 |
| (%) | 2 weeks | $202 \pm 24^{****}$ | $230 \pm 18^{****}$ |
| | 8 weeks | 136 ± 18** | $131 \pm 8*$ |

Table 2. Implant osseointegration assessed by micro-CT; bone volume per total tissue volume (BV/TV) and the mean trabecular number (Tb.N); *p < 0.05: HAPc group vs CG.

| Rat group | CG | HAPc |
|-------------|--------------|------------------|
| BV/TV(%) | 25.5±4.3 | $38.8 \pm 5.4^*$ |
| Tb.N (1/mm) | $154{\pm}18$ | $180{\pm}18^{*}$ |

Figure 3. Optical microscopy images of H&E stained slides of tissue samples at bone-implant interface near the fracture site, at eight weeks after implantation. (A): control group revealed fibrous tissue in proximity of Ti intramedullary implant and residual cartilaginous tissue indicating a transition from cartilaginous precursors to incipient bone trabeculae formation. (B): HAPc group displayed around HAPcTi implants the bone trabeculae well defined, compact bone with lamellar disposition of bone matrix and osteocytes around Havers canals, with osteoblasts lining their surface, and clear delimitation of areole between the trabeculae with areas of compact lamellar bone deposition.

Chapter 3 continues with the exploration of the *Behavior of multisubstituted hydroxyapatites inwater and simulated body fluid* (SBF). In order to determine the ion release profiles of these substituted hydroxyapatites [3]. Substituted nano-HAPs containg Mg, Zn, Sr and Si are synthesizes and characterized and it was demonstrated that substitution with essential elements in the HAP structure leads to the controlled release of the constituent ions from the HAP lattice, depending on the amount of doping element. While in water the release of the constituent ions is observed (Figure 6), in SBF this process is countered by the uptake by the sold HAP of Ca, Mg and Si from the solution (Figure 7). The long time release observed for the valuable physiological elements contained in multisubstituted hydroxyapatites evidences a promising future of these biomaterials for biomedical purposes.

Magnesium is found in bones and teeth. It is involved in the growth and remodeling of bones by activating osteoblast cells. Magnesium deficiency is linked to the onset of osteroporosis. *Zinc* inhibits osteoclasts and intensifies the response of osteoblasts. Anti-inflammatory and antimicrobial effects of zinc-substituted HAPs have also been reported. *Strontium* is known for its action on bone regeneration by increasing osteoblast activity and decreasing bone resorption by acting on osteoclasts. Strontium ranelate is increasingly used in the treatment of osteoporosis. *Silicon* is also involved in the bioactivity of osteoblasts.

For the synthesis of hydroxyapatites a solution containing the cations and one containing the anions were prepared. The 0.25 M cation solution contained Ca^{2 +} and, in addition, for complex HAPs:Mg²⁺, Zn²⁺ and Sr²⁺, according to the composition to be obtained. It was prepared by dissolving in ultrapure water the nitrates: $Ca(NO_3)_2 \cdot 4H_2O$, $Mg(NO_3)_2 \cdot 6H_2O$, $Zn(NO_3)_2 \cdot 6H_2O$ and $Sr(NO_3)_2$. The 0.15 M anions solution contained PO_4^{3-} and (for complex HAPs) SiO_4^{4-} . The solution was obtained from diammonium hydrogen phosphate, $(NH_4)2HPO_4$ and tetraethyl orthosilicate, TEOS, Si $(OC_2H_5)_4(98\%)$ in the appropriate ratio. The working pH was 11.5, fixed by adding a 25% ammonia solution. Equal volumes of the two solutions were mixed rapidly at room temperature, (22°C), using a peristaltic pump and a 'Y' type impact reactor for the two liquid flows. The suspension obtained was matured in two stages: at 22°C for 24 hours and at 70°C for another 24 hours, with intermittent stirring. The final precipitate was filtered and washed repeatedly with ultrapure water at room temperature until nitrates were removed. The drying process was carried out by lyophilization followed by the calcination step at 300°C for one hour and the sample as disintegrated in a ball mill to obtain a fine powder.

The presence of the HAP network as the unique crystalline phase was established by XRD and FTIR spectroscopy. The chemical composition was confirmed by SEM-EDX. TEM, SEM and AFM imaging showed the morphology of these biomaterials. The release of elements into water and simulated body fluid (SBF) was monitored over time, from 1 to 90 days, using inductively coupled plasma optical emission spectrometry (ICP-OES).

The amounts of Ca, P, Si and Mg in solutions were measured after immersion of 0.15 g of each sample in 15 ml of ultrapure water, respectively in Kokubo's simulated body fluid (SBF) and incubation at 37°C in separate closed flaskss for each sample/day. After 1, 3, 7, 14, 21, 30, 60 and 90 days, the supernatant (after centrifugation) was filtered. For calibration, standard multi-element solutions were prepared by diluting Merck IV multi-element stock solutions to 1000 mg/L. The Zn content in the aqueous phase was below the detection limit for all samples.

During the release of ions into water (Figure 6), for substituted hydroxyapatites, the amount of Ca released is higher than from pure HAP and very similar for all complex hydroxyapatites. The P / Ca ratio in solution is higher than in solid samples, which indicates an incongruent dissolution process. The amount of Mg passed into the aqueous phase is disproportionately high compared to its content in solid samples. It is lower for HAPc-Sr samples, denoting a stabilization of complex HAP by the simultaneous presence of Mg and Sr. Sr release, Figure 6d, increases slowly over time. It is higher for a higher Sr content in the solid sample. The Si content in solutions, Figure 6e, is almost constant over time after the first day. It is the highest for HAPc and decreases with increasing Sr content.

Figure 6. Calcium (a), phosphorus (b), magnesium (c), strontium (d), and silicon (e) release in water after immersion of HAPs samples for 1 – 90 days.

Figure 7. Calcium (a), phosphorus (b), magnesium (c), strontium (d), and silicon (e) contents in SBF after immersion of HAPs samples for 1 day to 90 days.

Chapter 4 presents the *Higuchi model applied to ions release from hydroxyapatites* [10] continues the investigation of ions release from tetrasubstituted HAPs with Mg, Zn, Sr and Si in water and SBF, in static and simulated dynamic conditions. The results of the experimental ion release determinations were theoretically interpreted by a diffusion-based Higuchi kinetic model. By representing the amounts of ions released as a function of the square root of time,

the applicability of the Higuchi equation was verified (e.g. Figure 2 for simulated dynamic conditions).

Figure 2. Cumulated ion release vs. time^{1/2} (days^{1/2}) for 7 days in simulated dynamic conditions from noncalcined and calcined samples of HAP, HAPc-5%Sr (HAPc-Sr5) and HAPc-10%Sr (HAPc-Sr10). Release in water of $Ca^{2+}(A)$, P (phosphate ions, B), $Mg^{2+}(C)$, $Sr^{2+}(D)$, and Sr^{2+} in SBF (E). Vertical bars represent the standard deviations of the measured values

The behavior of the synthesized stoichiometric hydroxyapatite (HAP): $Ca_{10}(PO_4)_6(OH)_2$ and two multisubstituted hydroxyapatites (ms-HAPs), both containing 1.5 wt% Mg, 0.2% Zn, 0.2% Si and different amounts of Sr: HAPc-5% Sr, respectively HAPc-10% Sr, when immersed in water and simulated body fluid

(SBF) has been recently researched [[1, 3, 4, 5, 10]. The theoretical formulas for ms-HAP materials are: $Ca_{8.76}Mg_{0.63}Zn_{0.03}Sr_{0.58}(PO_4)_{5.93}(SiO_4)_{0.07}(OH)_{1.93}$ for HAPc-5% Sr and $Ca_{8.12}Mg_{0.65}Zn_{0.03}Sr_{1.20}(PO_4)_{5.93}(SiO_4)_{0.07}(OH)_{1.93}$ for HAPc-10% Sr. The release of Ca^{2+} , Mg^{2+} , Sr^{2+} , as well as P (phosphate) in water and the variation of ion content in SBF in contact with submerged HAPs samples was measured using inductively coupled plasma emission spectrometry (ICP-OES). Zn^{2+} and silicate ions could not be detected in the solutions, as they were below the ICP-OES detection limit. A static method was applied, in which the HAPs samples were kept in the immersion liquid in closed flasks for different periods of time, from 1 to 90 days, and a simulated dynamic method, when the immersion liquid was changed daily with a fresh one, for 7 days.

The Higuchi model is based on Fick diffusion, so it should be applicable when the internal diffusion of ions from the bulk of the particle to the surrounding liquid is the rate-determining step, and the released species is evenly distributed in a homogeneous matrix. In this case, the amount released of a species should be proportional to the square root of time. But the regression lines for the linearization $Mt = f(t^{1/2})$ do not pass through the origin of the coordinates, in agreement with the finding that diffusion is not the main process in the early stages of ion release. Consequently, a modified form of the Hguchi equation was tried

$$M_t = a + Kt^{1/2}$$
 (2)

where M_t is the cumulative amount of ions released at time t, and K is a rate constant of ion release, which depends on both the characteristics of HAP nanoparticles and on the properties of the released species, but also on the nature of the immersion medium and temperature.

Equation (2) is applied satisfactorily for the release of ions under static conditions, most of the values of the determination coefficient r^2 are over 0.9. The Higuchi equation applies better to Ca and P, the major constituents of hydroxyapatites, best to unsubstituted HAP. The equation fails in case of Sr release from HAPc-10% Sr in SBF, where ion exchange processes can occur between Ca²⁺ or Mg²⁺ ions in the solution and Sr²⁺ ions in the solid sample. Higher r^2 values are observed for the last days of interaction between hydroxyapatite and water in most ions and samples, the linearity of the representations $Mt = M_t = f(t^{1/2})$ becoming more evident, as can be seen in Figure 2.

Considering the parameter K as a measure of the diffusion rate, it results that the diffusion of calcium and phosphorus takes place faster from substituted HAPs than from the unsubstituted, while for HAPc-10% Sr the rate is lower than for HAPc-5% Sr. This could be explained by the distortion of the crystal lattice by substituting the Ca^{2+} ion with other cations of different sizes, which favors internal diffusion. On the other hand, the values of a (y-intercepts) can be considered as an extrapolation of the amount of ions released at time 0 and a measure of the initial solubility. For Ca^{2+} they are also higher in substitut HAPs, which confirms

the increase in solubility by substitution in hydroxyapatite. The diffusion rate of the Mg^{2+} ion is much higher than that of Sr^{2+} and also above the value for Ca^{2+} . This may be due to the smaller size of the Mg^{2+} ions (ionic radius 86 pm) compared to Sr^{2+} (132 pm) and Ca^{2+} (114 pm), and therefore the higher mobility of the former. Higher strontium release in SBF than in water can be attributed to ion exchange with Ca^{2+} and Mg^{2+} ions in the SBF composition. Different release rates for different ions also determine a composition of the dissolved material differentfrom that of the initial solid, i.e. an incongruent dissolution of the hydroxyapatites. In simulated dynamic conditions, the linearity of the relationship (2) for days 1-7 is very good; all coefficients of determination are over 0.95, most of them over 0.99.

The results show the importance of diffusion in the release of ions from multisubstituted HAPs, while the calculated kinetic parameters reveal the peculiarities in the release of different ions.

Chapter 5 [6] investigates the Antibacterial activity of silver nanoparticles obtained by co-reduction with sodium citrateand tannic acid. Nanomaterials are developed which, added in coatings, have antimicrobial effects. For this purpose, silver nanoparticles were synthesized by a "green" method: co-reduction with sodium citrate and tannic acid, and their antimicrobial action on *Escherichia coli* cultures was investigated. This simple and fast one-pot synthesis led to AgNPs, with controlled size, from 30 to 10 nm, as shown by STEM and AFM images. The presence of elemental silver is evident from the UV-VIs spectra and from the EDX spectra and element distribution maps (Figure 5).

Co-reduction of silver nitrate with sodium citrate and tannic acid at various Ag/TSC/ TA molar ratios was done by heating to boiling the AgNO₃ solution and adding the calculated amount of TSC and TA mixture. The solutions were kept boiling under continuous stirring for 15 min. The colloidal silver solution obtained by complete reduction of Ag⁺ to Ag had a concentration of 1 mM Ag for the Ag-TSC-TA molar ratio of 1: 7: 2 and 0.25 mM for the molar ratios 1: 7: 0.2, 1: 3: 0.2 and 1: 20: 0.1. The Jasco UV / Vis V650 spectrophotometer was used for UV-VIS absorption spectrum measurements, in the wavelength range from 800 to 190 nm. STEM is a combination of the Hitachi HD-2700 scanning electron microscope (SEM) and the transmission electron microscope (TEM), which operates at a maximum acceleration voltage of 200 kV. STEM is equipped with an Energy Dispersive X-ray Spectrometer (EDS), which has two EDX detectors from Oxford Instruments. STEM-EDS equipment was also used for elemental EDX analysis.

In energy dispersive X-ray spectroscopy, the EDX spectra of the nanoparticles obtained on STEM images (as in the example shown in Figure 5a, b) the presence of Ag as well as of elements from the organic compounds is highlighted (Figure 5b). In Figure 5c-f, the correponding multicolor distribution maps for the individual elements (C, N, O, Ag) can be seen. The distribution maps confirm the presence of Ag mainly in the nanoparticles, while the other elements are distributed over the

entire scanned surface, due to the presence of organic compounds (TSC, TA and their oxidation products) around the particles and between them, being adsorbed on the STEM grid from the deposited colloidal solution.

Figure 5. Electronic image (a) and EDX spectrum (b), for the silver nanoparticle (in the white frame, shown in panel a), obtained for sample Ag: TSC: TA with a molar ratio of 1: 3: 0.2; distribution maps for C (c), N (d), O (e) and Ag (f); the bars in the images are 50 nm.

The results of antibacterial testing have shown that, in order to increase the antibacterial activity of AgNPs, various reaction parameters should be considered, including the molar ratio between reducing agent (TSC) and stabilizing (coating) agent, TA of the nanoparticle which seems to be a crucial factor, to finally produce stable nanoparticles of different sizes, preferably with small average diameters. Our results confirm the higher antibacterial activity of the smaller particles. This effect can be explained by the theory which assigns the antibacterial effect to Ag nanoparticles, because smaller particles have a larger specific surface area, being able to interact more strongly with the cell membrane or can penetrate the cell. On the other hand, if the antibacterial effect were due to Ag^+ ions, there is again the larger specific surface area the one that would guarantee a more intense release of silver ions. To these reasonings we could add that, for the diffusion in agar plates, which is slower than in liquid medium, the smaller particles have a higher mobility, thus increasing the inhibition zones.

Chapter 6 focuses on *the Interaction of silver nanoparticles withvancomycin: an UV-VIS study* [8]. To enhance the antibacterial effect of AgNP, but also of antibiotics, against which bacteria develop resistance, the possibility of using tigether AgNPs and antibiotics, for example vancomycin, was studied. For this purpose, the interaction with vancomycin of AgNPs prepared with different reducing and coating agents was thoroughly investigated: trisodium citrate, β -cyclodextrin, glucose-starch mixture (Figure3), glucose-TEOS (Figure 4), citric-tannic acid in various ratios. This knowledge of the characteristic behavior of vancomycin AgNP systems can help select the appropriate systems to maximize their antimicrobial effect.

For some preparations (e.g. AgNPs-citrate or AgNPs-citrate-tannicacid) stable AgNPs-vancomycin associations (complexes) were obtained. Forother colloidal solutions (e.g. AgNPs- β CD) less or more advanced vancomycinmediated self-assemblies of AgNPs appeared, and still maintained in colloidal solution. Finally, in other systems (e.g. AgNPs-glucose-starch or AgNPs-glucose-TEOS), the AgNPs slightly precipitated under the influence of vancomycin.

Figure 1. UV-VIS spectra of AgNPscitrateand V solutions in different ratios and in time

Figure 2. UV-VIS spectra of AgNPs- β -cyclodextrin (β -CD) and V solutions in different ratios and in time

Figure 3.UV-VIS spectraof AgNPsglucose-starch (gluc-starch) and V solutions in different ratios

Figure 5. UV-VIS spectra of AgNPscitrate (C) and tannicacid (T) in the molar ratio 1:7:2 and V solutions in different ratios and in time

Figure 4.UV-VIS spectra of AgNPsglucose-TEOS and V solutions and in time indifferent ratios and in time

Figure 6. UV-VIS spectra of AgNPscitrate (C) and tannic acid (T) in the molar ratio1:20:0.1 and V solutions in different ratios and in time

Chapter 7 aims to research *Biomimetic nanocomposite structures* designed for the coating of orthopedic implants: AFM investigation [11]. The complex coatings made on Ti implants, containing multisubstituted HAPs, were subjected to a detailed study by AFM, following all the stages of material deposition on the Ti surface, from the preparation of the metal surface, to the formation of collagen fibers on the porous composite (Figures 5, 8, 12). The AFM study was supplemented by XRD investigations (Figure 6).

Ti rods were tested by X-ray diffraction (XRD). The obtained pattern evidences the diffraction peaks only for titanium proving the highest purity of Ti rods.

The rounded titanium rods were flattened with a hydraulic press and cut into sticks with a 20 mm length Both sides of the sticks were grinded with P500 abrasive paper for 10 minutes to obtain a proper texture of the active surface. The grinding debris was removed by intense washing with bi-distilled water, followed by an ultrasound cleaning. After cleaning the rods were chemically activated for 30 min with orto-phosphoric acid to obtain a perfectly clean and degreased surface.

Figure 5. AFM images of HAPc-6%COL (core-shell) nanoparticles: a) topographic image, b) phase image, c) amplitude image, d) 3D image, and e) profile along the arrow in panel (a). Scanned area 1 μm x 1 μm; Ra 2.29 nm; Rq 2.89 nm.

Figure 6. XRD pattern for HAPc-6%COL (core-shell) nanoparticles

Figure 8. AFM images of pores network on HAPc-6%COL@PLA surface: a) topographic image, b) phase image, c) amplitude image, d) 3D image, and e) profile along the arrow in panel (a). Scanned area $20 \ \mu m \ge 20 \ \mu m$; Ra 289 nm; Rq 360 nm.

Figure 12. AFM images of the HAPc-6% COL@PLA/COLcoating on Ti implant: a) topographic image, b) phase image, c) amplitude image, d) 3D image, and e) profile along the arrow in panel (a). Scanned area 1 µm x 1 µm; Ra 16.2 nm; Rq 18.2 nm

The major component in ther coating material is the freeze dried ms-HAP/6%COL core-shell NPs. The HAPc NPs were investigated by AFM, and appear well individualized particles adsorbed on the Ti surface (Figure 5).

The rod coating dispersions contained HAPcfunctionalized with 6% collagenin a solution of PLA in dichloromethane. The fluidity of this dispersion is

increased by the addition of acetone. The titanium surface prepared as described above was coated with successive layers, using the dip-coating method. A layer of pure collagen was transferred by vertical adsorption for 5 seconds from a solution rich in COL at pH 12, resulting in the biomimetic structure HAPc-6%COL@PLA/COL on the Ti implant.

Porosity is an important requirement for a bone biomimetic structure to assure enough space for osteoblasts adhesion and proliferation. The pores network is a useful structure as observed in Figure 8, for HAPc-6%COL@PLA coating on Ti implant. These pores are of submicron size and are lastly generate by the slower evaporation of dichloromethane (DCM) under drying process of biocomposite.

On the surface of the HAPc-6%COL@PLA/COL coating, the AFM images show the formation of collagen fibers by self-assembly (Figure 12).

Besides qualitative characterization, AFM image processing allowed quantitative determinations, such as measuring surface roughness, an important feature for their physical and biological role (Tables 1, 2).

| Table 1 | . The | surface | roughness, | Ra and | l Rq | (RMS), | of Ti | impla | ints b | before (| coating | s, |
|----------|-------|---------|------------|--------|------|--------|-------|-------|--------|----------|---------|----|
| evaluate | ed by | AFM. | | | | | | | | | | |

| Ti | | Ti Ti | | Ti | | |
|------|--------|---------|---------|--------|-----------|------------|
| | Cold | pressed | Grinded | | Grinded a | and etched |
| | | _ | | | with | acid |
| Fig. | Ra±SD | RMS±SD | Ra±SD | RMS±SD | Ra±SD | RMS±SD |
| | nm | nm | nm | nm | nm | nm |
| 2 | 265±25 | 330±30 | - | - | - | - |
| 3 | - | - | 154±17 | 186±19 | - | - |
| 4 | - | - | | | 176±18 | 218±20 |

Table 2. The surface roughness, Ra and Rq (RMS), of biomimetic coatings on Ti implants measured by AFM

| Composite | HAPc-6% | COL@PLA | HAPc-6%CC | DL@PLA/COL |
|-----------|---------|---------|-----------|------------|
| Fig. | Ra±SD | RMS±SD | Ra±SD | RMS±SD |
| | nm | nm | nm | nm |
| 8 | 289±24 | 360±27 | - | - |
| 10 | - | - | 256±15 | 304±26 |

Chapter 8 GENERAL CONCLUSIONS

1). In the research of the doctoral thesis, we aimed to develop biomimetic composites for coating Ti implants (Chapter 1). The biomimetic coating (structure) was successfully developed using multisubstituted hydroxyapatite (ms-HAP) functionalized with collagen, COL (i.e. ms-HAP/COL nanoparticles core/shell), embedded in polylactic acid matrix (PLA), resulting in a porous biomimetic structure ms-HAP/COL@PLA, and subsequently coated with a self-assembled laver fibrous biomimetic of COL fibers. obtaining а composite ms-HAP/COL@PLA/COL, called HAPc. Subsequently, these implants were tested in vivo to assess bone consolidation in the absence or presence of high frequency pulsed electromagnetic waves (HF-PESW)

2). For *in vivo* evaluation, albino rats divided into four groups were used: control group (CG) with Ti implant; PESW group with Ti + HF-PESW implant; HAPc group with Ti implant coated with HAPc; HAPc + PESW group with Ti implant coated with HAPc + HF-PESW. The left femoral diaphysiswas fractured and fixed intramedullary. From the first postoperative day, the PESW and HAPc + PESW groups underwent HF-PESW stimulation for 14 consecutive days. The biomimetic coating was characterized by XRD, HR-TEM, SEM, EDX and AFM.

3) The use of HAPc-coated Ti implants together with HF-PESW stimulation positively influenced the bone consolidation process, especially in its early phase.

4). This *in vivo* evaluation demonstrated that the association between HF-PESW stimulation and HAPc coating on Ti implants promotes an accelerated healing process of bone fracture, enhancing bone consolidation in its early phase. Consequently, this combined method is potentially interesting and useful for clinical applications, proving a superior approach to the surface modification of biomedical implants.

5). The biocompatibility of uncovered titanium, Ti, nails, and coated with an innovative biocomposite was also assessed on a rat model of femoral fracture (**Chapter 2**). The biocomposite is based on multisubstituted hydroxyapatite, ms-HAP, containing Mg, Zn and Si, and is used as a coating material deposited on Ti implants, due to the excellent biocompatibility and osteoconductive property of ms-HAP.

6). Intramedullary titanium nails coated with multisubstituted hydroxyapatite and collagen in a polylactic acid matrix stimulate bone healing and also increased implant osseointegration into the host bone. In the case of clinical application of these implants, they could reduce the risk of implant default.

7). In order to clarify the contribution of multisubstituted hydroxyapatites to the delvery of ions with an important biochemical role, the behavior of multisubstituted hydroxyapatites in water and simulated body fluid, SBF, was studied (**Chapter 3**).

8).The introduction of essential elements with important biological effects in nanostructured hydroxyapatite has been demonstrated by physico-chemical investigations. Multisubstituted hydroxyapatites showed an average degree of crystallinity and a particle size in the nanometric range. The presence of Mg and Zn has a destabilizing effect on the HAP network, while the addition of Sr diminishes this effect.

9). The release of the component elements of these multisubstituted hydroxyapatites was examined in aqueous solutions, as a model for biological fluids. The increased concentration of Sr in the HAP structure significantly influenced the release of Sr in both environments: water and simulated body fluid.

10). A different profile for Ca, Mg and P is determined in both environments, and the formation of a new biomimetic hydroxyapatite is apparently promoted in SBF.

11). The sustained release observed for thevaluable physiological elements contained in multisubstituted hydroxyapatites reveals a promising future for these biomaterials for biomedical purposes.

12). The release of ions from multisubstituted hydroxyapatites was interpreted on the basis of a theoretical model (Higuchi model) in **Chapter 4**.

13). Considering the validity ranges of the Higuchi model for the ions release from the investigated HAPs, we can affirm that, while diffusion is important throughout the entire process of ion release in static conditions, from day 1 to 90, dissolution has also a significant contribution in the initial phase of the process. After the dissolution of the outer, more soluble, shell of particles, the internal diffusion of ions from the bulk to the interface with the immersion medium will be the main process. Moreover, in time a saturation of the solution is approached due to the low solubility, so diffusion remains predominant.

14). In simulated dynamic condition, when the immersion liquid is daily renewed, no saturation could occur, so both dissolution and diffusion contribute to the ion release.

15). To enhance the antimicrobial activity of hydroxyapatites, they can be combined (doped) with substances with antimicrobial action, such as silver nanoparticles or various antibiotics.

16). The characterization of silver nanoparticles obtained by co-reduction with sodium citrate and tannic acid was performed by UV-VS spectroscopy, scanning transmission electron microscopy (STEM) and theywere evaluated by atomic force

microscopy (AFM) images. Their antibacterial effect was highlighted against *Escherichia coli* (Chapter 5).

17). Co-reduction of silver nitrate in the aqueous solution with TSC and TA proved to be a simple and fast one-pot synthesis to prepare AgNPs, with controlled size, from 30 to 10 nm, as shown in STEM and AFM images. The presence of elemental silver is evident from the UV-VIS spectra (SPR characteristic band) and from the EDX spectra and element distribution maps. The antibacterial effect of AgNP was tested by measuring areas of inhibition on *Escherichia coli* cultures.

18). The effect was obvious for all AgNPs samples, but the dispersions with the smallest particle size proved to be the most active. Thus, AgNPs obtained by the investigated methods could be successfully used, as such or in combination with antibiotics, against bacterial infections.

19). The antimicrobial activity of antibiotics can be potentiated by using them together with silver nanoparticles. The interaction of such an antibiotic, vancomycin with silver nanoparticles was studied by the UV-VIS method (**Chapter 6**).

20). UV-VIS measurements allowed us to evence the different behavior of colloidal AgNPs solutions, obtained using different reducing systems, due to the formation of AgNPs with different coating (stabilization) agents. Furthermore, the interaction of these AgNPs with antibiotics, such as vancomycin, is identified.

21). For some preparations (e.g. AgNPs-citrate or AgNPs-citrate-tannicacid) stable AgNPs-vancomycin associations (complexes) were obtained. Forother colloidal solutions (e.g. AgNPs- β CD) less or more advanced vancomycinmediated self-assemblies of AgNPs appeared, and still maintained in colloidalsolution. Finally, in other systems (e.g. AgNPs-glucose-starch or AgNPs-glucose-TEOS), the AgNPs slightly precipitated under the influence of vancomycin.

22). The developing of biomimetic coating onto the Ti surface proves to be a smart choice to enhance the osseointegration and ensure an optimal healing process, due to the creation of nanostructured biomaterials similar to those in native bone. Thus, we designed a composite coating based on multi-substituted hydroxyapatite (noted ms-HAP or HAPc) nanoparticles, NPs, doped with essential elements: Mg, Zn and Si, functionalized with collagen type 1 (COL), embedded into poly lactic acid, PLA, matrix, and finally covered with COL layer to achieve biomimetic structures. Thin layers of biomimetic composite were self-assembled onto Ti surface via dip-coating method. Both, initial and coated Ti implants were investigated by atomic force microscopy (AFM), which allows surface investigation at high resolution of nano-level (**Chapter 7**).

23). AFM is a powerful tool for investigation of the biomimetic composite coating on the titanium surface of implants. It proves that the collagen amount in the nanocomposite material is able to reticulate, as COL fibers. The AFM images revealed a biomimetic network of collagen fibers similar to the one in natural bone formed on the surface of nano-composite layers.

24). The nano-topography and surface roughness are evidenced by AFM microscopy in the coating layers on Ti implants and are suitable for osteoblasts attachment to the surface increasing the cells viability and prolifertion.

25). Adding an extra layer of pure collagen on the coatig layers could be a model of enhancing the osteoblasts activity to generate new bone on the revealed biomimetic structures.

List of Research Papers- Original Contribution (Reka Balint)

8 Articles were published in scientific journals ISI, Cumulative impact factor:
8.515; 3 articles are accepted for publication (*in press*) in scientific journals ISI;
2 BDI articles and one article in Proceedings

1. P. T. Frangopol, A. Mocanu, V. Almasan, C. Garbo, **R. Balint**, G. Borodi, I. Bratu, O. Horovitz, M. Tomoaia-Cotisel, Synthesis and structural characterization of strontium substituted hydroxyapatites, Revue Roumaine de Chimie, 61(4-5), 337-344, (2016). IF = 0.304.

2. F. Goga, E. Forizs, G. Borodi, Gh. Tomoaia, A. Avram, **R. Balint**, A. Mocanu, O. Horovitz, M. Tomoaia Cotisel, Behavior of doped hydroxyapatites during the heat treatment, Rev. Chim.(Bucharest), 68(12), 2907-2913, (2017). IF = 1.412.

3. O. Cadar, **R. Balint**, Gh. Tomoaia, D. Florea, I. Petean, A. Mocanu, O. Horovitz, M. Tomoaia-Cotisel, Behavior of multisubstituted hydroxyapatites in water and simulated body fluid, Studia UBB Chemia, 62(4), Tom II, 269-281, (2017). IF = 0.305.

4. A. Mocanu, **R. Balint**, C. Garbo, L. Timis, I. Petean, O. Horovitz, M. Tomoaia-Cotisel , Low crystallinity nanohydroxyapatite prepared at room temperature, Studia UBB Chemia, 62(2), Tom I, 95-103, (2017). IF = 0.305.

5. A. Avram, M. Gorea, **R. Balint**, L. Timis, S. Jitaru, A. Mocanu, M. Tomoaia-Cotisel, Portland cement enriched with hydroxyapatite for endodontic applications, Studia UBB Chemia, 62(4), Tom I, 81-92, (2017). IF = 0.305.

6. S. Rapuntean, **R. Balint**, G. A. Paltinean, G. Tomoaia, A. Mocanu, C.-P. Racz, O. Horovitz, M. Tomoaia-Cotisel, Antibacterial activity of silver nanoparticles obtained by co-reduction with sodium citrate and tannic acid, Studia UBB Chemia, 63(3), 73-85, (2018). IF = 0.275.

7. D. Oltean-Dan, G.-B. Dogaru, M. Tomoaia-Cotisel, D. Apostu, A. Mester, H.-R.-C. Benea, M.-G. Paiusan, E.-M. Jianu, A. Mocanu, **R. Balint**, O.-C. Popa, C. Berce, G.-I. Bodizs, A.-M. Toader, Gh. Tomoaia, Enhancement of bone consolidation using high frequency pulsed electromagnetic short-waves and titanium implants coated with biomimetic composite embedded into PLA matrix: in vivo evaluation, Int J Nanomedicine, 14, 5799–5816, (2019). IF = 5.115.

8. **R. Balint**, G. A. Paltinean, A. Mocanu, O. Horovitz, M. Tomoaia-Cotisel, Interaction of silver nanoparticles with vancomycin: an UV-VIS study, Studia UBB Chemia, 64(2), Tom II, 335-343, (2019). IF = 0.494.

9. D. Oltean-Dan, P.T. Frangopol, **R. Balint**, Gh.Tomoaia, A. Mocanu, M. Tomoaia-Cotisel, Biocompatibility of titanium implants coated with biocomposite in a rat model of femoral fracture, Studia UBB Chemia, 66(3), (2021). *In press*.

10. A. Mocanu, P. T. Frangopol, **R. Balint**, O. Cadar, I. Vancea, R. Mantiu, O. Horovitz, M. Tomoaia-Cotisel, Higuchi model applied to ions release rate from hydroxyapatites, Studia UBB Chemia, 66(3), (2021). *In press*.

11. **R. Balint**, I. Petean, P.T. Frangopol, A. Mocanu, G. Arghir, O. Horovitz, M. Tomoaia-Cotisel, Biomimetic nanocomposite structures designed to cover orthopedic implants: AFM research, Studia UBB Chemia, 66(3), (2021). *In press.*

12. C. R. Popa, **R. Balint**, A. Mocanu, M. Tomoaia-Cotisel, Cardiovascular diseases induced by air pollution, Annals - Series on Biological Sciences, 9 (2), 133-170, (2020). ISSN 2285 – 4177.

13. **R. Balint**, G. A. Paltinean, G. Tomoaia, D. Oltean-Dan, A. Mocanu, M. Tomoaia-Cotisel, Review Biomaterials with enhanced biological functions for medical applications, Annals - Series on Biological Sciences, 10(1), 90 - 145, (2021). ISSN 2285 - 4177.

14. **R. Balint**, M. Tomoaia-Cotisel, Gh. Tomoaia, "Compozite biomimetice avansate utilizate în căptusirea implantelor metalice pentru vindecarea fracturilor osoase", Academia oamenilor de stiintă din România (AOSR), Conferinta de toamna, "Convergenta reala Romania-Uniunea Europeana, CRUE", Comunicari integrale, Vol. II, Brasov, 20-21 Septembrie, pages 266-274, (2019), ISBN 978-973-618-430-

Chapter 9. SELECTIVE REFERENCES

D. Oltean-Dan, G.-B. Dogaru, M. Tomoaia-Cotisel, D. Apostu, A. Mester, H.-R.-C. Benea, M.-G. Paiusan, E.-M. Jianu, A. Mocanu, **R. Balint**, O.-C. Popa, C. Berce, G.-I. Bodizs, A.-M. Toader, Gh. Tomoaia, Enhancement of bone consolidation using high frequency pulsed electromagnetic short-waves and titanium implants coated with biomimetic composite embedded into PLA matrix: In Vivo Evaluation, Int J Nanomedicine, 14, 5799–5816, (2019).

D. Oltean-Dan, P. T. Frangopol, **R. Balint**, Gh. Tomoaia, A. Mocanu, M. Tomoaia-Cotisel, Biocompatibility of titanium implants coated with biocomposite in a rat model of femoral fracture, Studia UBB Chemia, 66(3), (2021).

O. Cadar, **R. Balint**, Gh. Tomoaia, D. Florea, I. Petean, A. Mocanu, O. Horovitz, M. Tomoaia-Cotisel, Behavior of multisubstituted hydroxyapatites in water and simulated body fluid, Studia UBB Chemia, 62(4), Tom II, 269-281, (2017).

S. Rapuntean, **R. Balint**, G.A. Paltinean, G. Tomoaia, A. Mocanu, C.-P. Racz, O. Horovitz, M. Tomoaia-Cotisel, Antibacterial activity of silver nanoparticles obtained by co-reduction with sodium citrate and tannic acid, Studia UBB Chemia, 63(3), 73-85, (2018).

R. Balint, G. A. Paltinean, A. Mocanu, O. Horovitz, M. Tomoaia-Cotisel, Interaction of Silver Nanoparticles with Vancomycin: An UV-VIS Study, Studia UBB Chemia, 64(2), Tom II, 335-343, (2019).

R. Balint, I. Petean, P.T. Frangopol, A. Mocanu, G. Arghir, S. Riga, Gh. Tomoaia, O. Horovitz, M. Tomoaia-Cotisel, Structuri nanocompozite biomimetice proiectate pentru acoperirea implanturilor ortopedice: cercetare AFM, Studia UBB Chemia, 66(3), (2021).

A. Avram, M. Gorea, **R. Balint**, L. Timis, S. Jitaru, A. Mocanu, M. Tomoaia-Cotisel, Portland cement enriched with hydroxyapatite for endodontic Applications, Studia UBB Chemia, 62(4), Tom I, 81-92, (2017).

Gh. Tomoaia, R.D. Pasca, On the collagen mineralization. A review, Clujul Med, 88(1), 15–22, (2015). doi:10.15386/cjmed-359.

Gh. Tomoaia, O. Soritau, M. Tomoaia-Cotisel, L.-B. Pop, A. Pop, A. Mocanu, O. Horovitz, L.-D. Bobos, Scaffolds made of nanostructured phosphates, collagen and chitosan for cell culture, Powder Technol, 238, 99–107, (2013). doi:10.1016/j.powtec.2012.05.023.

Gh. Tomoaia, A. Mocanu, I. Vida-Simiti, N. Jumate, L.D. Bobos, O. Soritau, M. Tomoaia-Cotisel, Silicon effect on the composition and structure of nanocalcium phosphates: in vitro biocompatibility to human osteoblasts, Mater Sci Eng C Mater Biol Appl., 37(1), 37–47, (2014). doi:10.1016/j.msec.2013.12.027.

A. Mocanu, G. Furtos, S. Rapuntean, O. Horovitz, C. Flore, C. Garbo, A. Danistean, G. Rapuntean, C. Prejmerean, M. Tomoaia-Cotisel, Synthesis; characterization and antimicrobial effects of composites based on multi-substituted hydroxyapatite and silver nanoparticles, Appl Surf Sci., 298, 225–235, (2014). doi:10.1016/j.apsusc.2014.01.166.

F. Goga, E. Forizs, A. Avram, A. Rotaru, A. Lucian, I. Petean, A. Mocanu, M. Tomoaia-Cotisel, Synthesis and thermal treatment of hydroxyapatite doped with magnesium, zinc and silicon, Rev Chim., 68(6), 1193–1200, (2017).

F. Goga, E. Forizs, G. Borodi, Gh. Tomoaia, A. Avram, **R. Balint**, A. Mocanu, O. Horovitz, M. Tomoaia-Cotisel, Behavior of doped hydroxyapatites during the heat treatment, Rev Chim., 68(12), 2907–2913, (2017).

J.A. Buza, T. Einhorn, Bone healing in 2016, Clin Cases Miner Bone Metab, 13(2),101–105, (2016). doi:10.11138/ccmbm/2016.13.2.101.

E. Gómez-Barrena, P. Rosset, D. Lozano, J. Stanovici, C. Ermthaller, F. Gerbhard, Bone fracture healing: cell therapy in delayed unions and nonunions, Bone, 70, 93–101, (2015). doi:10.1016/j.bone.2014.07.033.

R. Dimitriou, G.I. Mataliotakis, A.G. Angoules, N.K. Kanakaris, P.V. Giannoudis ,Complications following autologous bone graft harvesting from the iliac crest and using the RIA: a systematic review, Injury, 42(suppl2), S3–S15, (2011). doi:10.1016/j.injury.2011.06.015.

J.W. Lu, F. Yang, Q.F. Ke, X.T. Xie, Y.P. Guo, Magnetic nanoparticles modified porous scaffolds for bone regeneration and photothermal therapy against tumors, Nanomedicine, 14(3), 811–822, (2018). doi:10.1016/j.nano.2017.12.025.

I. Antoniac, C. Sinescu, A. Antoniac, Adhesion aspects in biomaterials and medical devices, J Adhes Sci Technol, 30(16), 1711–1715, (2016).

J.V. Rau, I. Antoniac, G. Cama, V.S. Komlev, A. Ravaglioli, Bioactive materials for bone tissue engineering, Biomed Res Int, 2016 (Article ID 3741428), 1–3, (2016). doi:10.1155/2016/3741428.

F.G. Lyons, J.P. Gleeson, S. Partap, K. Coghlan, FJ. O'Brien, Novel microhydroxyapatite particles in a collagen scaffold: a bioactivebone void filler, Clin Orthop Relat Res., 472(4), 1318–1328, (2014). doi:10.1007/s11999-013-3438-0.

I. Degasne, M.F. Baslé, V. Demais, G. Huré, M. Lesourd, B. Grolleau, L. Mercier, D. Chappard, Effects of roughness, fibronectin and vitronectin on attachment, spreading, and proliferation of human osteoblast-like cells (Saos-2) on titanium surfaces, Calcif Tissue Int., 64(6), 499–507, (1999).

K.-H. Frosch, I. Sondergeld, K. Dresing, T. Rudy, C.H. Lohmann, J. Rabba, D. Schild, J. Breme, K.M. Stuerme, Autologous osteoblasts enhance osseointegration of porous titanium implants, J Orthop Res., 21(2), 213–223, (2003). doi:10.1016/S0736-0266(02)00143-2.

M. Shahrezaee, M. Salehi, S. Keshtkari, A. Oryan, A. Kamali, B. Shekarchi, In vitro and in vivo investigation of PLA/PCL scaffold coated with metformin-loaded gelatin nanocarriers in regeneration of critical-sized bone defects, Nanomedicine, 14(7), 2061–2073, (2018), doi:10.1016/j.nano.2018.06.007.

G.M. Cunniffe, C.M. Curtin, E.M. Thompson, G.R. Dickson, F.J. O'Brien, Content-dependent osteogenic response of nanohydroxyapatite: an invitro and in vivo assessment within collagen-based scaffolds, ACS Appl Mater Interfaces, 8(36), 23477–23488, (2016). doi:10.1021/acsami.6b06596.

F. Yu, M. Li, Z. Yuan, F. Rao, X. Fang, B. Jiang, Y. Wen, P. Zhang, Mechanism research on a bioactiveresveratrol–PLA–gelatin porous nano-scaffold in promoting therepair of cartilage defect, Int J Nanomed., 13, 7845–7858, (2018). doi:10.2147/IJN.S181855.

A. Tsuchiya, S. Sotome, Y. Asou, M. Kikuchi, Y. Koyama, T. Ogawa, J. Tanaka, K. Shinomiya, Effects of pore size and implantvolume of porous hydroxyapatite/collagen (HAp/Col) on bone formationin a rabbit bone defect model, J Med Dent Sci., 55(1), 91–99, (2008).

D.M. Ibrahim, A.A. Mostafa, S.I. Korowash, Chemical characterization of some substituted hydroxyapatites, Chem Cent J., 5(1), 74, (2011). doi:10.1186/1752-153X-5-74.

I. Izquierdo-Barba, L. Santos-Ruiz, J. Becerra, M.J. Feito, D. Fernández-Villa, M.C. Serrano, I. Díaz-Güemes, B. Fernández-Tomé, S. Enciso, F.M. Sánchez-Margallo, D. Monopoli, H. Afonso, M.T. Portolés, D. Arcos, M.Vallet-Regí, Synergistic effect of Si-hydroxyapatite coating and VEGF adsorption on Ti6Al4V-ELI scaffolds for bone regeneration in an osteoporotic bone environment, Acta Biomater, 83, 456–466, (2019). doi:10.1016/j.actbio.2018.11.017

A. Zhu, P. Lu, H. Wu, Immobilization of $poly(\varepsilon$ -caprolactone)– poly(ethylene oxide)–poly(ε -caprolactone) triblock copolymer on poly(lactide-coglycolide) surface and dual biofunctional effects, Appl Surf Sci., 253(6), 3247–3253, (2007). doi:10.1016/j.apsusc.2006.07.036.

J. Li, X.L. Lu, Y.F. Zheng, Effect of surface modified hydroxyapatite on the tensile property improvement of HA/PLA composite, Appl Surf Sci., 255(2), 494–497, (2008). doi:10.1016/j.apsusc.2008.06.067.

M. Hrubovcakova, M. Kupkova, M. Dzupon, M. Giretova, L. Medvecky, R. Dzunda, Biodegradable polylactic acid and polylactic acid/hydroxyapatite coated iron foams for bone replacement materials, Int J Electrochem Sci., 12(12), 11122–11136, (2017). doi:10.20964/2017.12.53.

W. Yang, G. Yin, D. Zhou, J. Gu, Y. Li, H. Zhang, Biocompatibility of surface-modified biphasic calcium phosphate/poly-L-lactide biocomposite vitro and in vivo, J Mater Sci Technol., 26(8), 754–758, (2010). doi:10.1016/S1005-0302(10)60119-3.

P. Vashisth, J.R. Bellare, Development of hybrid scaffold with biomimetic 3D architecture for bone regeneration, Nanomed., 14(4), 1325–1336, (2018). doi:10.1016/j.nano.2018.03.011.

Y. Shapovalova, D. Lytkina, L. Rasskazova, A. Gudima, V. Ryabov, A. Filimoshkin, I. Kurzina, J. Kzhyshkowska, P97: bioresorbable composites based on hydroxyapatite dispersed in poly-L-lactidematrix, Eur J Cancer Suppl, 13(1), 49–50, (2015). doi:10.1016/j.ejcsup.2015.08.088.

P. Kubasiewicz-Ross, J. Hadzik, J. Seeliger, K. Kozak, K.l Jurczyszyn, H. Gerber, M. Dominiak, C.Kunert-Keil, New nano-hydroxyapatite in bone defect regeneration: a histological study in rats, Ann Anat., 213, 83–90, (2017). doi:10.1016/j.aanat.2017.05.010.

F. Ren, Y. Leng, R. Xin, X. Ge, Synthesis, characterization and initio simulation of magnesium-substituted hydroxyapatite, Acta Biomater., 6(7), 2787–2796, (2010). doi:10.1016/j.actbio.2009.12.044.

P.T. Frangopol, A. Mocanu, V. Almasan, C Garbo. R. Balint, G. Borodi, I. Bratu, O. Horovitz, M. Tomoaia-Cotisel, Synthesis and structural characterization of strontium substituted hydroxyapatites, Rev Roum Chim.,61(4–5), 337–344, (2016).

Gh. Tomoaia, L.B. Pop, I. Petean, M. Tomoaia-Cotisel, Significance of surface structure on orthopedic materials, Mater Plast., 49(1), 48–54, (2012).

Gh. Tomoaia, M. Tomoaia-Cotisel, L.B. Pop, A. Pop, O. Horovitz, A. Mocanu, N. Jumate, L.D. Bobos, Synthesis and characterization of some composites based on nanostructured phosphates, collagen and chitosan, Rev Roum Chim., 56(10–11), 1039–1046, (2011).

J. Russias, E. Saiz, R.K. Nalla, K. Gryn, R.O. Ritchie, A.P. Tomsia, Fabrication and mechanical properties of PLA/HA composites: a study of in vitro degradation, Mater Sci Eng C Biomim Supramol Syst., 26(8), 1289–1295, (2006). doi:10.1016/j.msec.2005.08.004.

M.T. Tsai, W.J. Li, R.S. Tuan, W.H. Chang, Modulation of osteogenesis in human mesenchymal stem cells by specific pulsed electromagnetic field stimulation, J Orthop Res., 27(9), 1169–1174, (2009). doi:10.1002/jor.20862.

K. Chang, W.H. Chang, S. Huang, S. Huang, C. Shih, Pulsed electromagnetic fields stimulation affects osteoclast formation by modulation of osteoprotegerin, RANK ligand and macrophage colony-stimulating factor, J Orthop Res., 23(6), 1308–1314, (2005). doi:10.1016/j.orthres.2005.03.012.1100230611.

V. Galkowski, P. Brad, B. Drew, D. Dick, Bone stimulation for fracture healing: what's all the fuss?, Indian J Orthop, 43(2), 117–120, (2009). doi:10.4103/0019-5413.50844.

K.F. Taylor, N. Inoue, B. Rafiee, J.E. Tis, K.A. McHale, E.Y. Chao, Effect of pulsed electromagnetic fields on maturation of regenerate bone in arabbit limb lengthening model, J Orthop Res., 24(1), 2–10, (2006). doi:10.1002/jor.20014.

P.F. Hannemann, E.H. Mommers, J.P. Schots, P.R. Brink, M. Poeze, The effects of low-intensity pulsed ultrasound and pulsed electromagnetic fields bone growth stimulation in acute fractures: a systematic review and meta analysis of randomized controlled trials, Arch Orthop Trauma Surg., 134(8), 1093–1106, (2014). doi:10.1007/s00402-014-2014-8.

C.M. Teven, M. Greives, R.B. Natale, Y. Su, Q. Luo, B.-C. He, D. Shenaq, T.-C. He, R.R. Reid, Differentiation of osteoprogenitor cells is induced by high-frequency pulsed electromagnetic fields. J Craniofac Surg., 23(2), 586–593, (2012). doi:10.1097/SCS.0b013e31824cd6de.

Gh. Tomoaia, M. Tomoaia-Cotisel, A. Mocanu, O. Horovitz, L.D. Bobos, M. Crisan, I. Petean, Supramolecular organization of collagen and anti-cancer drugs, J Optoelectron Adv Mater., 10(4), 961–964, (2008).

A. Mocanu, R.D. Pasca, Gh. Tomoaia, C. Garbo, P.T. Franghopol, O. Horovitz, M. Tomoaia-Cotisel, New procedure to synthesize silver nanoparticles and their interaction with local anesthetics, Int J Nanomedicine, 8, 3867–3874, (2013). doi:10.2147/IJN.S51063.

J.W. Park, Y.J. Kim, J.H. Jang, T.G. Kwon, Y.C. Bae, J Y. Suh, Effects of phosphoric acid treatment of titanium surfaces on surface properties, osteoblast response and removal of torque forces, Acta Biomater., 6(4), 1661–1670, (2010). doi:10.1016/j.actbio.2009.10.011.

D. Jing, M. Zhai, S. Tong, F. Su, J. Cai, G. Shen, Y. Wu, X. Li, K.Xie, J. Liu, Q. Xu, E. Luo, Pulsed electromagnetic fields promote osteogenesis and osseointegration of porous titanium implants in bone defect repair through a Wnt/β-catenin signaling-associated mechanism, Sci Rep., 6(1), 32045, (2016). doi:10.1038/srep32045.

J.V. Rau, M. Fosca, I. Cacciotti, S. Laureti, A. Bianco, R. Teghil, Nanostructured Si-substituted hydroxyapatite coatings for biomedical applications, Thin Solid Films, 543, 167–170, (2013). doi:10.1016/j.tsf.2012.12.113.

F. Loi, L.A. Cordova, J. Pajarinen, T.H. Lin, Z. Yao, S.B. Goodman Inflammation, fracture and bone repair, Bone, 86, 119–130. (2016). doi:10.1016/j.bone.2016.02.020.

W. Lin, L. Xu, S. Zwingenberger, E. Gibon, S.B. Goodman, G. Li, Mesenchymal stem cells homing to improve bone healing, J Orthop Transl, 9, 19– 27, (2017).

S. Mirhadi, N. Ashwood, B. Karagkevrekis, Factors influencing fracture healing. Trauma., 15(2), 140–155, (2013). doi:10.1177/1460408613486571.

D.R. Marsh, G. Li, The biology of fracture healing: optimising outcome, Br Med Bull., 55(4),856–869, (1999). doi:10.1258/0007142991902673.

A. Klymov, J. Te Riet, P. Mulder, J.G.E. Gardeniers, J.A. Jansen, X.F. Walboomers, Nanometer-grooved topography stimulates trabecular bone regeneration around a concave implant in a rat femoral medulla model, Nanomed, 12(8), 2283–2290, (2016). doi:10.1016/j.nano.2016.06.013.

T. Wehner, L. Claes, A. Ignatius, U. Simon, Optimization of intramedullary nailing by numerical simulation of fracture healing, J Orthop Res., 30(4),569–573, (2012). doi:10.1002/jor.21568.

M. Goutam, C. Giriyapura, S.K. Mishra, S. Gupta, Titanium allergy: a literature review, Indian J Dermatol., 59(6), 630, (2014). doi:10.4103/0019-5154.143526.

D. Crouzier, L. Selek, B.A. Martz, V. Dabouis, R. Arnaud, J.C. Debouzy, Risk assessment of electromagnetic fields exposure with metallic orthopedic implants: a cadaveric study, Orthop Traumatol Surg Res., 98(1), 90–96, (2012). doi:10.1016/j.otsr.2011.08.012.

E.P. Buzza, J.A. Shibli, R.H. Barbeiro, J.R. Barbosa, Effects of electromagnetic field on bone healing around commercially pure titanium surface: histologic and mechanical study in rabbits, Implant Dent., 12(2),182–187, (2003).

Z. Zhang, Z. Li, C. Zhang, J. Liu, Y. Bai, S. Li, C. Zhang, Biomimetic intrafibrillar mineralized collagen promotes bone regeneration via activation of the Wnt signaling pathway, Int J Nanomed., 13, 7503–7516, (2018). doi:10.2147/IJN.S172164.

H.T. Sirin, I. Vargel, T. Kutsal, P. Korkusuz, E. Piskin, Ti implants with nanostructured and HA-coated surfaces for improved osseointegration, Artif Cells Nanomedi Biotechnol., 44(3), 1023–1030, (2016). doi:10.3109/21691401.2015.1008512.

X. Lin, S. Yang, K. Lai, H. Yang, T.J. Webster, L. Yang, Orthopedic implant biomaterials with both osteogenic and anti-infection capacities and

associated in vivo evaluation methods, Nanomed, 13(1), 123–142, (2017).doi:10.1016/j.nano.2016.08.003.

N. Attia, M. Mashal, S. Grijalvo, R. Eritja, J. Zárate, G. Puras, J. L.Pedraz, Stem cell-based gene delivery mediated by cationic niosomes for bone regeneration, Nanomedicine, 14(2), 521–531, (2018). doi:10.1016/j.nano.2017.11.005.

J. Henkel, M.A. Woodruff, D.R. Epari, R. Steck, V. Glatt, I.C. Dickinson, P. F. M. Choong, M.A. Schuetz, D.W. Hutmacher, Bone regeneration based on tissue engineering conceptions – a 21st century perspective, Bone Res., 1(3), 216–248, (2013). doi:10.4248/BR201303002.

Q. Ding, X. Zhang, Y. Huang, Y. Yan, X. Pang, In vitro cytocompatibility and corrosion resistance of zinc-doped hydroxyapatite coatings on a titanium substrate, J Mater Sci., 50(1), 189–202, (2015). doi:10.1007/s10853-014-8578-4.

C. Garbo, M. Sindilaru, A. Carlea, Gh. Tomoaia, V. Almasan, I. Petean, A. Mocanu, O. Horovitz, Synthesis and structural characterization of novel porous zinc substituted nanohydroxyapatite powders, Part Sci Technol., 35(1), 29–37, (2017). doi:10.1080/02726351.2015.1121180.

Y.L. Cai, J. Zhang, S. Zhang, S.S. Venkatraman, X.T. Zeng, H. J. Du, D. Mondal, Osteoblastic cell response on fluoridated hydroxyapatite coatings: the effect of magnesium incorporation, Biomed Mater., 5(5), 054114, (2010), doi:10.1088/1748-6041/5/3/035008.

E.S. Thian, J. Huang, S.M. Best, Z.H. Barber, W. Bonfield, Novel silicon doped hydroxyapatite (Si-HA) for biomedical coatings: an in vitro study using a cellular simulated body fluid, J Biomed Mater Res B Appl Biomater., 76B(2), 326–333, (2006). doi:10.1002/jbm.b.30368.

I.J. Macha, B. Ben-Nissan, J. Santos, S. Cazalbou, A. Stamboulis, D. Grossin, G. Giordano, Biocompatibility of a new biodegradable polymerhydroxyapatite composite for biomedical applications, J Drug Deliv Sci Technol, 38, 72–77, (2017). doi:10.1016/j.jddst.2017.01.008.

G.R. Beck, S.W. Ha, C.E. Camalier, M. Yamaguchi, Y. Li, J. –K. Lee, M. N. Weitzmann, Bioactive silica-based nanoparticles stimulate bone-forming osteoblasts, suppress bone-resorbing osteoclasts, and enhance bone mineral density in vivo, Nanomed, 8(6), 793–803, (2012). doi:10.1016/j.nano.2011.11.003.

M. Uezono, K. Takakuda, M. Kikuchi, S. Suzuki, K. Moriyama, Hydroxyapatite/collagen nanocomposite-coated titanium rod for achieving rapid osseointegration onto bone surface, J Biomed Mater Res B Appl Biomater, 101B(6), 1031–1038, (2013). doi:10.1002/jbm.b.32913.

A. Murakami, T. Arimoto, D. Suzuki, M. Iwai-Yoshida, F. Otsuka, Y. Shibata, T. Igarashi, R. Kamijo, T. Miyazaki, Antimicrobial and osteogenic properties of a hydrophilic-modified nanoscale hydroxyapatite coating on titanium, Nanomed, 8(3), 374–382, (2012). doi:10.1016/j.nano.2011.07.001.

S. Itoh, M. Kikuchi, Y. Koyama, H. N. Matumoto, K. Takakuda, K. Shinomiya, J. Tanaka, Development of a novel biomaterial, hydroxyapatite/collagen (HAp/Col) composite for medical use, Biomed Mater Eng., 15(1–2), 29–41, (2005).

X. Wei, S. Egawa, R. Matsumoto, H.Yasuda, K. Hirai, T. Yoshii, A. Okawa, T. Nakajima, S. Sotome, Augmentation of fracture healing by hydroxyapatite/collagen paste and bone morphogenetic protein-2 evaluated using a

rat femur osteotomy model, J Orthop Res., 36(1), 129–137, (2018). doi:10.1002/jor.23646.

G.G. Walmsley, A. McArdle, R. Tevlin, Arash Momeni , D. Atashroo, M. S. Hu, A.H. Feroze, V.W. Wong, P.H. Lorenz, M.T. Longaker, D.C. Wan, Nanotechnology in bone tissue engineering, Nanomedicine, 11(5), 1253–1263, (2015). doi:10.1016/j.nano.2015.02.013.

S. Sprio, A. Tampieri, E. Landi, M. Sandri, S. Martorana, G. Celotti, G. Logroscino, Physico-chemical properties and solubility behaviour of multisubstituted hydroxyapatite powders containing silicon, Mater Sci Eng C., 28(1), 179–187, (2008). doi:10.1016/j.msec.2006.11.009.

D. Govindaraj, M. Rajan, M.A. Munusamy, A.A. Alarfai, K.K. Sadasivuni, S.S. Kumar, The synthesis, characterization and in vivo study of mineral substituted hydroxyapatite for prospective bone tissue rejuvenation applications, Nanomedicine, 13(8), 2661–2669, (2017). doi:10.1016/j.nano.2017.07.017.

L. Kyllönen, M. D'Este, M. Alini, D. Eglin, Local drug delivery for enhancing fracture healing in osteoporotic bone, Acta Biomater., 11(1), 412–434, (2015). doi:10.1016/j.actbio.2014.09.006.

F. Veronesi, M. Cadossi, G. Giavaresi, L. Martini, S. Setti, R. Buda, S. Giannini, M. Fini, Pulsed electromagnetic fields combined with a collagenous scaffold and bone marrow concentrate enhance osteochondral regeneration: an in vivo study, BMC Musculoskelet Disord., 16(1), 233, (2015).

A.F. Mavrogenis, R. Dimitriou, J. Parvizi, G. C. Babis, Biology of implant osseointegration, J. Musculoskelet, 9(2), 61-71, (2009).

P. I. Brånemark, Osseointegration and its experimental background, J. Prosthet. Dent., 50(3), 399-410, (1983).

R. D. Bloebaum, K. N. Bachus, N. G. Momberger, A. A. Hofmann, Mineral apposition rates of human cancellous bone at the interface of porous coated implants, J. Biomed. Mater. Res., 28(5), 537–544, (1994).

R. Depprich, H. Zipprich, M. Ommerborn, C. Naujoks, H. P. Wiesmann, S. Kiattavorncharoen, H. C. Lauer, U. Meyer, N. R. Kubler, J. Handschel, Osseointegration of zirconia implants compared with titanium: an in vivo study, Head Face Med., 4, 30, (2008).DOI: 10.1186/1746-160X-4-30.

P. Thomsen, C. Larsson, L. E. Ericson, L. Sennerby, J. Lausmaa, B. Kasemo, Structure of the interface between rabbit cortical bone and implants of gold, zirconium and titanium, J. Mater. Sci. Mater. Med., 8(11), 653–665, (1997).

A. B. Novaes Jr, S. L. Scombatti de Souza, R. R. Martins de Barros, K. K. Y. Pereira, G. Iezzi, A. Piattelli, Influence of implant surfaces on osseointegration, Braz. Dent. J., 21(6), 471–481, (2010).

W. Wang, C.K. Poh, Titanium Alloys in Orthopaedics, In Titanium Alloys - Advances in Properties Control, J. Sieniawski, W. Ziaja Eds.; Intech Open; London, UK, 2013, chapter 1, pp. 1-20.

J. T. B. Ratnayake, M. Mucalo, G. J. Dias, Substituted hydroxyapatites for bone regeneration: A review of current trends, J. Biomed. Mater. Res. B Appl. Biomater., 105(5), 1285–1299, (2017). D. Shepherd, S. M. Best,Production of zinc substituted hydroxyapatite using various precipitation routes, Biomed. Mater., 8(2), 025003, (2013). DOI: 10.1088/1748-6041/8/2/025003.

A. Tsuchiya, S. Sotome, Y. Asou, M. Kikuchi, Y. Koyama, T. Ogawa, J. Tanaka, K. Shinomiya, Effects of pore size and implant volume of porous hydroxyapatite/collagen (HAp/Col) on bone formation in a rabbit bone defect model, J. Med. Dent. Sci., 55(1), 91-99, (2008).

D. Govindaraj, M. Rajan, M. A. Munusamy, A. A. Alarfaj, K. K. Sadasivuni, S. S. Kumar, The synthesis, characterization and in vivo study of mineral substituted hydroxyapatite for prospective bone tissue rejuvenation applications, Nanomed, 13(8), 2661-2669, (2017).

Gh. Tomoaia, M. Tomoaia-Cotisel, A. Mocanu, O. Horovitz, L. D. Bobos, M. Crisan, I. Petean, Characterization by atomic force microscopy of some composites based on surface active glasses and copolymers, J. Optoelectron. Adv. Mater., 10(4), 961-964, (2008).

F. Yang, W. J. Dong, F. M. He, X. X. Wang, S. F. Zhao, G. L. Yang, Osteoblast response to porous titanium surfaces coated with zinc-substituted hydroxyapatite, Oral Surg. Oral Med. Oral Pathol. Oral Radiol., 113(3), 313-318, (2012).

S. Rapuntean, P.T. Frangopol, I. Hodisan, Gh. Tomoaia, D. Oltean-Dan, A. Mocanu, C. Prejmerean, O. Soritau, L.Z. Racz, M. Tomoaia-Cotisel, In vitro response of human osteoblasts cultured on strontium substituted hydroxyapatites, Rev Chim (Bucharest), 69(12), 3537-3544, (2018).

C. Garbo, J. Locs, M. D'Este, G. Demazeau, A. Mocanu, C. Roman, O. Horovitz, M. Tomoaia-Cotisel, Advanced Mg, Zn, Sr, Si multi-substituted hydroxyapatites for bone regeneration, Int. J. Nanomed., 15, 1037-1058, (2020).

A. Mocanu, O. Cadar, P. T. Frangopol, I. Petean, Gh. Tomoaia, G. A. Paltinean, Cs. P. Racz, O. Horovitz, M. Tomoaia-Cotisel, Ion release from hydroxyapatite and substituted hydroxyapatites in different immersion liquids: in vitro experiments and theoretical modelling study, R. Soc. Open. Sci., 8(1), 201785, (2021).

E. Boanini, M. Gazzano, A. Bigi, Ionic substitutions in calcium phosphates synthesized at low temperature, Acta Biomater, 6(6), 1882-1894, (2010).

E.S. Thian, J. Huang, M. E. Vickers, S. M. Best, Z.H. Barber, W. Bonfield, Silicon-substituted hydroxyapatite (SiHA): A novel calcium phosphate coating for biomedical applications, J. Mater. Sci., 41(3), 709-717, (2006).

A. Mocanu, **R. Balint**, C. Garbo, L. Timis, I. Petean, O. Horovitz, M. Tomoaia- Cotisel, Low crystallinity nanohydroxyapatite prepared at room temperature, Studia UBB Chemia, 62(2), Tom I, 95-103, (2017).

J. Beuvelot, Y. Mauras, G. Mabilleau, H. Marchand-Libouban, D. Chapparda, Adsorption and release of strontium from hydroxyapatite crystals developed in simulated body fluid (SBF) on poly (2-hydroxyethyl) methacrylate substrates, Dig. J. Nanomater. Biostructures, 8(1), 207-217, (2013).

M. Tomoaia-Cotisel, N. Cioica, C. Cota, Cs. Racz, I. Petean, L. D. Bobos, A. Mocanu, O. Horovitz, Structure of starch granules revealed by atomic force microscopy, Studia UBB Chemia, 55 (2), Tom II, 313-326 (2010).

R.D. Pasca, Gh. Tomoaia, A. Mocanu, I. Petean, G.A. Paltinean, O. Soritau, M. Tomoaia-Cotisel, Porous Collagen Scaffolds for Bone Regeneration, Studia UBB Chemia, 60(3), 257-264, (2015).

M.A. Naghiu, M. Gorea, E. Mutch, F. Kristaly, M. Tomoaia-Cotisel, Forsterite nanopowder: structural characterization and biocompatibility evaluation, Mater. Sci. Technol., 29(7), 628-632, (2013).

A. Danistean, M. Gorea, A. Avram, S. Rapuntean, Gh. Tomoaia, A. Mocanu, C. Garbo, O. Horovitz, M. Tomoaia-Cotisel, Antimicrobial activity of ceramic disks loaded with silver ions and nitroxoline, Studia UBB Chemia, 61 (3), 275-283, (2016).

Gh. Tomoaia, O. Horovitz, A. Mocanu, A. Nita, A. Avram, C.P. Racz, O. Soritau, M. Cenariu, M. Tomoaia-Cotisel, Effects of doxorubicin mediated by gold nanoparticles and resveratrol in two human cervical tumor cell lines, Colloids Surf. B, 135, 726-734, (2015).

P.T. Frangopol. D.A. Cadenhead, Gh. Tomoaia, A. Mocanu, M. Tomoaia-Cotisel, The effect of procaine on lipid domains investigated by contact mode atomic force microscopy, Rev. Roum. Chim, 60(2-3), 265-273, (2015).

G. Furtos, M.A. Naghiu, H. Declercq, M. Gorea, C. Prejmerean, O. Pana, M. Tomoaia-Cotisel, Nano forsterite biocomposites for medical applications: Mechanical properties and bioactivity, J. Biomed. Mater. Res. Part B Appl. Biomater. 104(7), 1290-1301, (2016).

M. Tomoaia-Cotisel, A. Mocanu, Phase transitions in phospholipid monolayers studied by atomic force microscopy and Langmuir-Blodgett technique, Rev. Chim. (Bucharest), 59, 1230-1233, (2008).

O. Horovitz, Gh. Tomoaia, A. Mocanu, T. Yupsanis, M. Tomoaia-Cotisel, Protein binding to gold autoassembled films, Gold Bull, 40 (4), 295-304, (2007).

M. Tomoaia-Cotisel, A. Tomoaia-Cotisel, T. Yupsanis, G. Tomoaia, I. Balea, A. Mocanu, Cs. Racz, Coating layers of major storage protein from aleurone cells of barley studied by atomic force microscopy, Rev. Roum. Chim, 51 (12), 1181-1185, (2006).

Gh. Tomoaia, A. Mocanu, L.D. Bobos, L.B. Pop, O. Horovitz, M. Tomoaia-Cotisel, Biocomposites for Orthopedic Applications, Studia UBB Chemia, 60 (3), 265-272, (2015).

T. Kokubo, H. Kushitani, S. Sakka, T. Kitsugi, T. Yamamuro, Solutions able to reproduce in vivo surface-structure changes in bioactive glass-ceramic A-W, J. Biomed. Mater. Res., 24(6), 721-734, (1990).

E. Forizs, F. Goga, A. Avram, A. Mocanu, I. Petean, O. Horovitz, M. Tomoaia-Cotisel, Thermal analysis of pure and multisubstituted hydroxyapatite pastes, Studia UBB Chemia, 62 (4, Tom I), 173-180, (2017).

Gh. Tomoaia, M. Tomoaia-Cotisel, L.B. Pop, A. Mocanu, A. Pop, Nanopowders of hydroxyapatite and its substituted derivatives with medical applications and their fabrication procedure, Romanian Patent, OSIM, Bucharest, Romania, no. 125817; BOPI, 6, 123, (2013).

M. Rohnke, S. Pfitzenreuter, B. Mogwitz, A. Henß, J. Thomas, D. Bieberstein, T.Gemming, S. K. Otto, S. Ray, M. Schumacher, M. Gelinsky, V. Alt,

Strontium release from Sr^{2+} -loaded bone cements and dispersion in healthy and osteoporotic rat bone, J. Control. Release, 262,159–169, (2017).

U. Gbureck, E .Vorndran, J.E. Barralet, Modeling vancomycin release kinetics from microporous calcium phosphate ceramics comparing static and dynamic immersion conditions, Acta Biomater., 4(5), 1480–1486, (2008).

A. Mocanu, O. Cadar, P.T. Frangopol, I. Petean, Gh. Tomoaia, G.A. Paltinean, C.P. Racz, O. Horovitz, M. Tomoaia-Cotisel, Ion release from hydroxyapatite and substituted hydroxyapatites in different immersion liquids: in vitro experiments and theoretical modelling study, Roy. Soc. Open Sci. 8, 201785, (2021).

T. Higuchi, Mechanism of sustained-action medication. Theoretical analysis of rate of release of solid drugs dispersed in solid matrices, J. Pharm. Sci., 52, 1145–1149, (1963).

S. Dash, P. N. Murthy, L. Nath, P. Chowdhury, Kinetic modeling on drug release from controlled drug delivery systems, Acta Pol. Pharm., 67, 217-223, (2010).

J. Siepmann, N.A. Peppas, Higuchi equation: Derivation, applications, use and misuse, Int J Pharm., 418, 6-12, (2011).

H.-G. Lee, Y.-S. Park, J.-H. Jeong, Y.-B. Kwon, D. H. Shin, J.-Y. Kim, Y.-S. Rhee, E.-S. Park, D.-W. Kim, C.-W. Park, Physicochemical properties and drugrelease mechanisms of dual-release bilayer tablet containing mirabegron and fesoterodine fumarate, Drug Des., Devel. Ther., 13, 2459–2474, (2019).

S. Bose, S. Tarafder, Calcium phosphate ceramic systems in growth factor and drug delivery for bone tissue engineering: A review, Acta Biomater., 8(4), 1401-1421, (2012).

M. M. Mailafiya,K. Abubakar, A. Danmaigoro, S. M. Chiroma, E. B. A. Rahim, M. A. M. Moklas, Z.A.B. Zakaria, Evaluation of In-Vitro Release Kinetic and Mechanisms of Curcumin Loaded-Cockle Shell-Derived Calcium Carbonate Nanoparticles, Biomed. Res. Ther., 6, 3518-3540, (2019).

E. Landi, A. Tampieri, G. Celotti, S. Sprio, M. Sandri, G. Logroscino, Srsubstituted hydroxyapatites for osteoporotic bone replacement, Acta Biomater., 3(6), 961-969, (2007).

S. V. Dorozhkin, Surface Reactions of Apatite Dissolution, J. Colloid Interf. Sci., 191(2), 489-497, (1997).

T. Kokubo, Biomaterials, Bioactive glass ceramics: properties and applications, Biomaterials, 12(2), 155-163, (1991).

J.R. Morones, J.L. Elechiguerra, A. Camacho, K. Holt, J.B. Kouri, J.T. Ramirez, M. J. Yacaman, The bactericidal effect of silver nanoparticles, Nanotechnology, 16(10), 2346-2353, (2005).

M. Rai, A. Yadav, A. Gade, Silver nanoparticles as a new generation of antimicrobials, Biotechnol. Adv., 27(1), 76-83, (2009).

M.K. Rai, S.D. Deshmukh, A.P. Ingle, A.K. Gade, Silver nanoparticles: the powerful nanoweapon against multidrug-resistant bacteria, J. Appl. Microbiol., 112(5), 841-852, (2012).

M.J. Hajipour, K.M. Fromm, A.A. Ashkarran, D.J. de Aberasturi, I.R. deLarramendi, T. Rojo, V. Serpooshan, W.J. Parak, M. Mahmoudi, Antibacterial properties of nanoparticles, Trends Biotechnol, 30(10), 499-511, (2012).

N. Durán, G. Nakazato, A.B. Seabra, Antimicrobial activity of biogenic silver nanoparticles, and silver chloride nanoparticles: an overview and comments, Appl. Microbiol. Biotechnol, 100(15), 6555-6570, (2016).

K.S. Siddiqi, A. Husen, R.A.K. Rao, A review on biosynthesis of silver nanoparticles and their biocidal properties, J. Nanobiotechnology, 16, 14, (2018). https://doi.org/10.1186/s12951-018-0334-5.

V. Chahar, B. Sharma, G. Shukla, A. Srivastava, A. Bhatnagar, Study of antimicrobial activity of silver nanoparticles synthesized using green and chemical approach A Physicochemical and engineering aspects, Colloids Surf, A Physicochem Eng Asp, 554, 149, (2018).

A. Mocanu, O. Horovitz, C.P. Racz, M. Tomoaia-Cotisel, Green synthesis and characterization of gold and silver nanoparticles, Rev. Roum. Chim, 60(7-8), 721-72, (2015).

Y. Hao, N. Zhang, J. Luo, X. Liu, Green Synthesis of Silver Nanoparticles by Tannic Acid with Improved Catalytic Performance Towards the Reduction of Methylene Blue, NANO: Brief Reports and Reviews, 13, 1850003, (2018). DOI: 10.1142/S1793292018500030.

P. Orlowski, M. Krzyzowska, R. Zdanowski, A. Winnicka, J. Nowakowska, W.Stankiewicz, E. Tomaszewska, G. Celichowski, J. Grobelny, Assessment of in vitro cellular responses of monocytes and keratinocytes to tannic acid modified silver nanoparticles, Toxicol. In Vitro, 27(6), 1798-1808, (2013).

S.K. Sivaraman, I. Elango, S. Kumar, V. Santhanam, A green protocol for room temperature synthesis of silver nanoparticles in seconds, Curr. Sci., 97(7), 1055-1059, (2009).

Y. Cao, R. Zheng, X Ji, H. Liu, R. Xie, W. Yang, Syntheses and characterization of nearly monodispersed, size-tunable silver nanoparticles over a wide size range of 7-200 nm by tannic acid reduction, Langmuir, 30(13), 3876-3882, (2014).

N.G. Bastus, F. Merkoci, J. Piella, V. Puntes, Synthesis of highly monodisperse citrate-stabilized silver nanoparticles of up to 200 nm: kinetic control and catalytic properties, Chem. Mater., 26, 2836-2846, (2014).

S. Agnihotri, S. Mukherji, S. Mukherji, Size-controlled silver nanoparticles synthesized over the range 5–100 nm using the same protocol and their antibacterial efficacy, RSC Advances, 4, 3974-3983, (2014).

O. Horovitz, M. Tomoaia-Cotisel, C. Racz, Gh. Tomoaia, L.D. Bobos, A. Mocanu, The interaction of silver nanoparticles with lipoic acid, Studia UBB Chemia, 54(3), 89-96, (2009).

Gh. Tomoaia, P.T. Frangopol, O. Horovitz, L.D. Bobos, A. Mocanu, M. Tomoaia-Cotisel, The effect of arginine on gold nanoparticles in colloidal solutions and in thin films, J. Nanosci. Nanotechnol, 11(9), 7762-7770, (2011).

U.V. Zdrenghea, Gh. Tomoaia, D.V Pop-Toader, A. Mocanu, O. Horovitz, M.Tomoaia-Cotisel, Procaine effect on human erythrocyte membrane explored by atomic force microscopy, Comb. Chem. High Throughput Screen., 14(4), 237-247, (2011).

R.D. Pasca, A. Mocanu, S.C. Cobzac, I. Petean, O. Horovitz, M. Tomoaia-Cotisel, Biogenic syntheses of gold nanoparticles using plant extracts, Particul Sci Technol, 32(2), 131-137, (2014).

I. Petean, G. Tomoaia, O. Horovitz, A. Mocanu, M. Tomoaia-Cotisel, Cysteine mediated assembly of gold nanoparticles, J. Optoelectron. Adv. Mater. J Optoelectron Adv M, 10(9), 2289-2292, (2008).

S. U. Khan, T. A Saleh, A. Wahab, M. H. U. Khan, D. Khan, W. U. Khan, A. Rahim, S. Kamal, F. U. Khan, S. Fahad, Nanosilver: new ageless and versatile biomedical therapeutic scaffold, Int. J. Nanomed, 13, 733-762, (2018).

L. Ge, Q. Li, M. Wang, J. Ouyang, X. Li, M. M. Q. Xing, Nanosilver particles in medical applications: synthesis, performance, and toxicity, Int. J. Nanomed, 9, 2399–2407, (2014).

A. Coates, Y. Hu, R. Bax, C. Page, The future challenges facing the development of new antimicrobial drugs, Nat. Rev. Drug Discov., 1, 895–910, (2002).

Y. E. Hur, Y. Park, Vancomycin-Functionalized Gold and Silver Nanoparticles as an Antibacterial Nanoplatform Against Methicillin-Resistant Staphylococcus aureus, J. Nanosci. Nanotechnol., 16, 6393-6399, (2016).

N. Xu, H. Cheng, J. Xu, F. Li, B. Gao, Z. Li, C. Gao, K. Huo, J. Fu, W. Xiong, Silver-loaded nanotubular structures enhanced bactericidal efficiency of antibiotics with synergistic effect in vitro and in vivo, Int. J. Nanomed, 12, 731-743, (2017).

A. Kaur, D. Goyal, R. Kumar, Surfactant mediated interaction of vancomycin with silver nanoparticles, Appl. Surf. Sci., 449, 23-30, (2018).

A. Kaur, S. Preet, V. Kumar, R. Kumar, R. Kumar, Synergetic effect of vancomycin loaded silver nanoparticles for enhanced antibacterial activity, Colloids Surf. B: Biointerfaces, 176, 62–69, (2019).

G. Furtos, M. Tomoaia-Cotisel, C. Garbo, M. Senila, N. Jumate, I. Vida-Simiti, C. Prejmerean, New composite bone cement based on hydroxyapatite and nanosilver, Particul. Sci. Technol., 31(4), 392-398, (2013).

S. Ahmed, M. Ahmad, B. L. Swami, S. Ikram, A review on plants extract mediated synthesis of silver nanoparticles for antimicrobial applications: A green expertise, J. Adv. Res. 7, 17–28, (2016).

T. A. Abalkhil, S. A. Alharbi, S. H. Salmen, M. Wainwright, Bactericidal activity of biosynthesized silver nanoparticles against human pathogenic bacteria, Biotechnol. Biotechnol. Equip., 31, 411–417, (2017).

S. Z. H. Naqvi, U. Kiran, M. I. Ali, A. Jamal, A. Hameed, S. Ahmed, N. Ali, Combined efficacy of biologically synthesized silver nanoparticles and different

antibiotics against multidrug-resistant bacteria, Int. J. Nanomed, 8, 3187–3195, (2013).

M. Tomoaia-Cotisel, Multifunctional nanostructures formed of gold or silver nanoparticles and different biomolecules with medical applications, e-Book, Cluj University Press, Cluj-Napoca, pp. 1-322, (2016). http://www.editura.ubbcluj.ro/bd/ebooks/pdf/1976.pdf

B. P. Howden, J. K. Davies, P. D. R. Johnson, T. P. Stinear, M. L. Grayson, Reduced Vancomycin Susceptibility in Staphylococcus aureus, Including Vancomycin-Intermediate and Heterogeneous Vancomycin-Intermediate Strains: Resistance Mechanisms, Laboratory Detection, and Clinical Implications, Clin. Microbiol. Rev., 23, 99-139, (2010).

M. Esmaeillou, G. Zarrini, M. A. Rezaee, J. S. Mojarrad, A. Bahadori, Vancomycin Capped with Silver Nanoparticles as an Antibacterial Agent against Multi-Drug Resistance Bacteria, Adv. Pharm. Bull., 7, 479-483, (2017).

I. Cacciotti, Multisubstituted hydroxyapatite powders and coatings: The influence of the codoping on the hydroxyapatite performances, Int J Appl Ceram Technol., 16(5), 1864-1884, (2019).

M.H. Santos, P. Valerio, A.M. Goes, M.F. Leite, L.G.D. Heneine, H.S. Mansur, Biocompatibility evaluation of hydroxyapatite/collagen nanocomposite doped with Zn^{+2} , Biomed. Mater. 2(2), 135–141, (2007).

I.V. Antoniac, A. Antoniac, E. Vasile, C. Tecu, M. Fosca, V. G. Yankova, J.V. Rau, In vitro characterization of novel nanostructured collagen-hydroxyapatite composite scaffolds doped with magnesium with improved biodegradation rate for hard tissue regeneration, Bioact. Mater., 6(10), 3383–3395, (2021).

I. Ullah, M. A. Siddiqui, S. K. Kolawole, H.Liu, J. Zhang, L. Ren, K. Yang, Synthesis, characterization and in vitro evaluation of zinc and strontium binary doped hydroxyapatite for biomedical application, Cerami Int, 46(10), 14448–14459, (2020).

T. Kumai, N. Yui, K. Yatabe, C. Sasaki, R. Fujii, M. Takenaga, H. Fujiya, H. Niki, K. Yudoh, A novel, self-assembled artificial cartilagehydroxyapatite conjugate for combined articular cartilage and subchondral bone repair: histopathological analysis of cartilage tissue engineering in rat knee joints, Int J Nanomed, 14, 1283–1298, (2019).

H. Liu, M. Lin, X. Liu, Y. Zhang, Y. Luo, Y. Pang, H. Chen, D. Zhu, X. Zhong, S. Ma, Y. Zhao, Q. Yang, X. Zhang, Doping bioactive elements into a collagen scaffold based on synchronous self-assembly/mineralization for bone tissue engineering, Bioact. Mater., 5(4), 844–858, (2020).

T. G. Kim, S.-H. Park, H. Jung Chung, D.-Y. Yang, T. G. Park, Microstructured scaffold coated with hydroxyapatite/collagen nanocomposite multilayer for enhanced osteogenic induction of human mesenchymal stem cells, J. Mater. Chem., 20, 8927–8933, (2010).

P. Pan, X. Chen, K. Metavarayuth, J. Su, Q. Wang, Self-assembled supramolecular systems for bone engineering applications, Curr. Opin. Colloid Interface Sci, 35, 104–111, (2018).

Z. Wang, Y. Yan, T. Wan, Fabrication and characterization of hydroxyapatite/collagen bone-like nanocomposite through a self-assembly method, Sci Eng Compos Mater, 19(2), 177–182, (2012).

A. Cappella, H.H. de Boer, P. Cammilli, D. De Angelis, C. Messina, L M. Sconfienza, F. Sardanelli, C. Sforza, C. Cattaneo, Histologic and radiological analysis on bone fractures: Estimation of posttraumatic survival time in skeletal trauma, Forensic Sci. Int., 302, 1-9, (2019).

E.N. L'Abbé, S.A. Symes, D.E. Raymond, D.H. Ubelaker, The Rorschach butterfly, understanding bone biomechanics prior to using nomenclature in bone trauma interpretations, Forensic Sci. Int., 299, 187 – 189, (2019).

I. Izquierdo-Barba, L. Santos-Ruiz, J. Becerra, M.J. Feito, D. Fernandez-Villa, M.C. Serrano, I. Diaz-Guemes, B. Fernandes Tome, S. Enciso, F.M. Sanches Margallo, D. Monopoli, H. Alfonso, M.T. Portoles, D. Arcos, M. Vallet-Regi, Synergistic effect of Si-hydroxyapatite coating and VEGF adsorption on Ti6Al4V-ELI scaffolds for bone regeneration in an osteoporotic bone environment, Acta Biomater., 83, 456–466, (2019).

L. Barbu-Tudoran, Gh. Tomoaia, O. Horovitz, A. Mocanuand M. Tomoaia-Cotisel, Self-assembly characteristics of gold nanoparticles in the presence of arginine, J. Optoelectron. Adv. Mater. J, 10(9), 2293-2297, (2008).

M. Tomoaia-Cotisel, A. Sen, P. J. Quinn, Surface active properties of 1,2distearoylgalactosylglycerols, J Colloid Interface Sci, 94(2), 390-398, (1983).

M. Tomoaia-Cotisel, J. Zsako, E. Chifu, Ejection curves and miscibility of egg lecithin with some carotenoid derivatives, Rev Roum Chim, 32(7), 663-670 (1987).

M. Tomoaia-Cotisel, J. Zsako, E. Chifu, P. J. Quinn, Intermolecular interactions in lipid-carotenoid monolayers, Biochem J, 248, 877-882, (1987).

M. Tomoaia-Cotisel, J. Zsako, E. Chifu, P. J. Quinn, Hysteresis in compression-expansion cycles of distearoylmonogalactosylglycerol monolayers, Chem Phys Lipids, 50, 127-133, (1989).

M. Tomoaia-Cotisel, J. Zsako, E. Chifu, D. A. Cadenhead, Relaxation phenomena in apocarotenoid monolayers, Langmuir, 6(1), 191-197 (1990).

M. Tomoaia-Cotisel, P. J. Quinn, Biophysical properties of carotenoids, in Subcellular Biochemistry, Fat-Soluble Vitamins, Editors: P. J. Quinn and V. E. Kagan, (Plenum Press, New York, USA, 1998), Vol. 30, Chapter 10, pp. 219-242. ISBN: 0-306-45846-2.

M. Tomoaia-Cotisel, T. Oproiu, J. Zsako, A. Mocanu, P. T. Frangopol, P. J. Quinn, Numerical analysis of compression isotherms of distearoyl monogalactosyl glycerol monolayers, Rev Roum Chim, 45(9), 851-861, (2000).

M. Tomoaia-Cotisel, E. Chifu, J. Zsako, P. T. Frangopol, P. J. Quinn, A. Mocanu, Interaction of some drugs with monomolecular membranes at the fluid interfaces, Studia UBB Chemia, 38(1-2), 81-85, (1993).

E. Chifu, M. Tomoaia-Cotisel, Insoluble monolayers of lecithin and carotenoid pigments, Rev Roum Chim, 24(7), 979-986, (1979).

E. Chifu, M. Tomoaia-Cotisel, Z. Andrei, Mixed monolayers of canthaxanthin with lipids, Studia UBB Chemia, 24(2), 63-67, (1979).

J. Zsako, M. Tomoaia-Cotisel, A. Mocanu, E. Chifu, Insoluble mixed monolayers. II. Protolytic equilibria and the influence of the pH on the collapse pressure, J Colloid Interface Sci, 110(2), 317-334, (1986).

E. Chifu, M. Tomoaia-Cotisel, Z. Andrei, E. Bonciu, β -apo-8-carotenoic acid ethyl ester films at fluid interfaces, Gazz Chim Ital, 109(6-7), 365-369, (1979).

M. Tomoaia-Cotisel, E. Chifu, J. Zsako, Mixed monolayers of egg lecithin and carotenoids, Colloids Surf, 14(2), 239-246, (1985).

M. Tomoaia-Cotisel, E. Chifu, Mixed insoluble monolayers with β -apo-8-carotenoic acid ethyl ester, Gazz Chim Ital, 109(6-7), 371-375, (1979).

E. Chifu, M. Tomoaia-Cotisel, A. Ioanette, Mixed insoluble monolayers of cholesterol and β -apo-8-carotenal, Gazz Chim Ital, 109(6-7), 397-398, (1979).

M. Tomoaia-Cotisel, E. Chifu, Carotenoid pigment films at fluid interface, Rev Chim (Bucharest), 32(11), 1063-1069 (1981).

I. Cojocaru, A. Tomoaia-Cotisel, A. Mocanu, T. Yupsanis, M. Tomoaia-Cotisel, The effect of globular protein from aleurone cells of barley on stearic acid monolayers, Rev Chim (Bucharest), 68(7), 1470-1475, (2017).

J. Zsako, M. Tomoaia-Cotisel, E. Chifu, A. Mocanu, P. T. Frangopol, Influence of stearic acid monolayers upon the procaine adsorption from underlying alkaline aqueous solutions, Biochim. Biophys. Acta, 1024(2), 227-232, (1990).

Chapter 10. ORIGINAL SCIENTIFIC RESEARCH

List of Original Research; Original Contributions- Reka BALINT (pag. 25)

8 Articles were published in scientific journals ISI, Cumulative impact factor: 8.515; 3 articles are accepted for publication in scientific journals ISI (*in press*); 2 BDI articles and **one** article in Proceedings

Total citations: 60, h-index 5; Google Scholar profile:

https://scholar.google.com/citations?user=SkRatYcAAAAJ&hl=ro

List of communications at conferences and symposia

The scientific results were presented at 13 conferences

1. C. Garbo, A. Mocanu, V. Almasan, **R. Balint**, G. Borodi, I. Bratu. O. Horovitz, M. Tomoaia-Cotisel, Effect of preparation conditions and the presence of surfactant on the hydroxyapatite nanopowders, PIM 2015, September 23-25, 2015

2. A. Danistean, C. Garbo, **R. Balint**, G. Tomoaia, A. Mocanu, S. Rapuntean, O. Horovitz, M. Tomoaia-Cotisel, Bioceramics comprising silver nanoparticles, as a

new generation of antimicrobial, Conferences on Physics of Advanced Materials, Cluj-Napoca, 8-10 September, 2016

3. **R. Balint**, C. Garbo, A. Mocanu, G. Tomoaia, I. Petean, O. Horovitz, M. Tomoaia-Cotisel, Revolving research discoveries into health, COST Action MP1301, Cluj-Napoca, 13-15 May, 2017

4. **R. Balint**, G. Tomoaia, S. Rapuntean, A. Mocanu, O. Horovitz and M. Tomoaia-Cotisel, Novel composites based on nanoceramics and silver nanoparticles with antimicrobial activity for biomedical applications, International Conference on Materials Science & Materials Chemistry, Paris, France, 20-22 August 2018 **ORAL PRESENTATION + POSTER**

5. D. Oltean-Dan, D. Apostu, G. B. Dogaru, G. Tomoaia, M. Tomoaia-Cotisel, A. Mester, M.-G. Paiusan, A. Mocanu, R. Balint, C. O. Popa, C. Berce, B. Gyorgy-Istvan, A. Toader, H.-R.-C. Benea, High frequency pulsed electromagnetic shortwaves and titanium nails coated with multi-substituted hydroxyapatite functionalized with collagen embedded intro PLA martri facilitates bone consolidation: an experimenta study, Abstract Volume of 8th International "Biomaterials. Devices" Conference Tissue Engineering and Medica (Biommedd' 2018) Cluj-Napoca, 27-29 September, 2018 ORAL PRESENTATION

6. **R. Balint,** M. Tomoaia-Cotisel, G. Tomoaia, "Compozite biomimetice avansate utilizate în căptușirea implantelor metalice pentru vindecarea fracturilor osoase", (Advanced biomimetic compositesused in coating metal implants for healing bone fractures) Conferință națională științifică Academia Oamenilor de Știința din Romănia, Book of abstract, **13**(2), 2019, **20-21 Septembrie Brasov, 80-81, ISSN 2601-5102 ORAL PRESENTATION**

7. **R Balint,** S Rapuntean, A Mocanu, O Horovitz and M Tomoaia-Cotisel,New antibacterial systems for biomedical applications,12th International Conference Processes in Isotopes and Molecules" (PIM), 25-27 September, 2019, Cluj-Napoca, Romania. **POSTER**

8. G. A. Paltinean, **R. Balint**, A. Mocanu, Gh. Rapuntean, I. Petean, O. Horovitz and M. Tomoaia-Cotisel, Antimicrobial activity of poly lactic acid microspheres loaded with vancomicin, 12th International Conference, Processes in Isotopes and Molecules (PIM), 25-27, September2019, Cluj-Napoca, Romania **POSTER**

9. Gh. Tomoaia, **R. Balint**, A. Mocanu,M. Tomoaia-Cotisel,Captuseli/straturi bioactive pe implante din titan (Bioactive coatings/layers ontitanium implants), Conferinta Nationala Stiintifica Academia Oamenilor de Stiinta din Romania, Secție Științe Biologice, 28 May-7 June 2020, **online**

10. Gh. Tomoaia, **R. Balint**, A. Mocanu, M. Tomoaia-Cotisel, Căptuşeli/straturi bioactive pe implante din titan (Bioactive coatings/layers ontitanium implants),, Conferinta Nationala Stiintifica Academia Oamenilor de Stiinta din Romania, Secție Științe Medicale, 18 iunie 2020, **online**

11.Gh. Tomoaia, **R. Balint**, A. Mocanu,M. Tomoaia-Cotisel, Materiale inovative pentru substitut de os (Innovative materials for bone substitutes),Conferinta Nationala Stiintifica Academia Oamenilor de Stiinta din Romania,Sesiunea Științifică de Toamna, Secție Științe Biologice, 27-28 November 2020, online

12. **R. Balint,** Gh. Tomoaia, D. Oltean-Dan, A. Mocanu, G. Arghir, M. Tomoaia-Cotisel,Nanomateriale cu funcție biologică îmbunătățită pentru regenerare osoasă (Nanomaterials with improved biological function for bone regeneration), Conferinta Nationala Stiintifica Academia Oamenilor de Stiinta din Romania, Secție Științe Biologice, 11 June 2021, **online**

13. **R. Balint**, Gh. Tomoaia, D. Oltean-Dan, A. Mocanu, M. Tomoaia-Cotisel, Biomaterials based on multifunctional hydroxyapatite for orthopedic applications, International Scientific Conference, Applications of chemistry in nanosciences and biomaterials engineering, Virtual Conference, 25 - 26 June 2021

Activity in research grants

The doctoral student has worked as a research assistant in **6 scientific research projects** carried out in the Center for Scientific Research in Physical Chemistry, CECHIF, under the supervision of Univ. Prof. Dr. Maria Tomoaia-Cotişel, at Faculty of Chemistry and Chemical Engineering, Babeş-Bolyai University of Cluj-Napoca.

List of scientific grants

1. PN2 Grant Partnerships 241/2014-2016

- 2. Grant Exploratory Research Project: PCE 83/2017-2019
- 3. Grant Experimental Demonstrative PNIII: Partnerships 481/2020-2022
- 4. Exploratory Research Project: PCE186/2021-2023

5. Mobility program for researchers.PN-III-P1-1.1-MC-2018-1737

Novel composites based on nanoceramics and silver nanoparticles with antimicrobial activity for biomedical applications; Participation at International Conference on Materials Science and Materials Chemistry 20-22 August, Paris, France. **Responsible for the project: Reka Balint.** **6. Reka Balint:** Advanced biomimetic composites awarded in 2019 by the Academy of Romanian Scientists.

15.4. Member in the Center of Scientific Research in Physical Chemistry

The doctoral student is a member of the Center of Scientific Research in Physical Chemistry, CECHIF, directed by the Director, Univ. Prof. Dr. Maria Tomoaia-Cotisel, Faculty of Chemistry and Chemical Engineering, Babeş-Bolyai University of Cluj-Napoca.